



Citation	Janssens Lizanne, Dinh Van Khuong, Stoks Robby, 2014, Extreme temperatures in the adult stage shape delayed effects of larval pesticide stress: a comparison between latitudes Aquatic Toxicology vol:148 pages:74-82.
Archived version	Author manuscript: the content is identical to the content of the published paper, but without the final typesetting by the publisher
Published version	http://dx.doi.org/10.1016/j.aquatox.2014.01.002
Journal homepage	http://www.journals.elsevier.com/aquatic-toxicology/
Author contact	your email lizanne.janssens@bio.kuleuven.be your phone number + 32 (0)16 32 38 57
IR	https://lirias.kuleuven.be/handle/123456789/448138

3

4

5

6

7

8

9

10

11

12

13

14

Extreme temperatures in the adult stage shape delayed effects of larval pesticide stress: a comparison between latitudes

Lizanne Janssens¹, Khuong Dinh Van¹, Robby Stoks¹

¹ Laboratory of Aquatic Ecology, Evolution and Conservation, University of Leuven, Charles Deberiotstraat 32, B-3000 Leuven, Belgium

Corresponding author: Lizanne Janssens

Tel. +3216323857; fax: +3216324575

E-mail addresses: lizanne.janssens@bio.kuleuven.be, khuong.dinhvan@bio.kuleuven.be, robby.stoks@bio.kuleuven.be

Abstract

Global warming and pesticide pollution are major threats for aquatic biodiversity. Yet, how pesticide effects are influenced by the increased frequency of extreme temperatures under global warming and how local thermal adaptation may mitigate these effects is unknown. We therefore investigated the combined impact of larval chlorpyrifos exposure, larval food stress and adult heat exposure on a set of fitness-related traits in replicated low- and high-latitude populations of the damselfly *Ischnura elegans*. Larval pesticide exposure resulted in lighter adults with a higher water content, lower fat content, higher Hsp70 levels and a lower immune function (PO activity). Heat exposure reduced water content, mass, fat content and flying ability. Importantly, both stressors interacted across metamorphosis: adult heat exposure lowered the reduction of fat content, and generated a stronger decrease in PO activity in pesticide-exposed animals. Larval pesticide exposure and larval food stress also reduced the defense response to the adult heat stress in terms of increased Hsp70 levels. In line with strong life history differences in the unstressed control situation, high-latitude animals were less sensitive to food stress (body mass and water content), but more sensitive to pesticide stress (development time and PO activity) and heat exposure (PO activity and Hsp70 levels). While low-latitude adults could better withstand the extreme temperature as suggested by the weaker increase in Hsp70, heat exposure similarly affected the delayed effects of larval pesticide exposure at both latitudes. Our study highlighted two key findings relevant for ecological risk assessment under global warming. Firstly, the delayed effects of larval pesticide exposure on adult damselflies depended upon subsequent adult heat exposure, indicating that larval pesticide stress and adult heat stress interacted across metamorphosis. Secondly, low- and high-latitude animals responded differently to the imposed stressors, highlighting that intraspecific evolution along natural thermal gradients may shape sensitivity to pesticides.

Keywords: carry-over effects; chlorpyrifos; damselfly; heat; latitudinal gradient; multiple stressors.

Introduction

High temperatures are receiving increased attention in ecotoxicology as these typically increase the toxicity of contaminants such as pesticides (Noyes et al., 2009) and therefore are critical when extrapolating toxicity tests across seasons and along thermal gradients (Clements et al., 2012). Especially the need to understand the impact of pesticides under global warming generated a surge of studies (Landis et al., 2013; Moe et al., 2013). These studies generated very valuable information for the development of a predictive framework for ecological risk assessment along natural thermal gradients and under global warming.

Studies on global warming typically exposed organisms to a moderate temperature increase, as predicted by relevant warming scenarios (IPCC, 2007). Such moderate temperature increases are often beneficial for temperate organisms in the absence of contaminants (Deutsch et al., 2008; Stoks et al., in prep.). Global warming will, however, also be characterized by more frequent temperature extremes (IPCC, 2012) and these are more likely to negatively impact organisms (Jentsch et al., 2007). Yet, temperature extremes received much less attention in ecotoxicology (but see Kaur et al., 2011).

While most studies simultaneously exposed organisms to contaminants and warming, there is increasing concern that stressors may interact in a delayed way (Segner, 2011). Such delayed effects are easily overlooked in the many animals that have a complex life cycle with a larval aquatic stage and an adult, terrestrial stage, separated by metamorphosis (Moran, 1994). Several studies have documented carry-over effects of larval pesticide exposure on adult traits (e.g. Campero et al., 2008; Distel and Boone, 2010; Boone et al. 2013), but few

have explored how exposure to contaminants in the larval stage affects the sensitivity of the animals to a different stressor in the adult stage (e.g. Rohr and Palmer, 2005) and no studies so far did this in a global warming framework and tested how delayed effects of larval pesticide exposure are shaped by adult exposure to extreme temperatures.

Another aspect that may further increase realism to assess the impact of contaminants under global warming is the consideration of thermal adaptation shaping the sensitivity to contaminants at higher temperatures (Moe et al., 2013). Given that temperature extremes are more frequent at lower latitudes (Orlowsky and Seneviratne, 2012) it is to be expected that low-latitude populations may suffer less from extreme temperatures (Hoffmann et al., 2002). This generates the hypothesis that temperature extremes will differentially affect the carry-over effects of larval pesticide stress in low- and high-latitude populations.

We addressed all three above-mentioned issues by investigating the combined impact of larval sublethal pesticide exposure and subsequent adult exposure to extreme temperatures on a set of fitness-related traits in a common garden experiment with replicated populations at the lower and higher part of the latitudinal range of an aquatic insect. Given their limited ability to escape exposure, many ectothermic aquatic organisms are particularly vulnerable to combinations of contaminants and high temperatures (Bronmark and Hansson, 2002; Woodward et al., 2010). We additionally manipulated food stress in the larval stage, because this may make energy-mediated carry-over effects more apparent (Karl et al., 2011). Moreover, suboptimal food conditions may increase the impact of pesticide stress (Janssens and Stoks, 2013a) and of exposure to extreme temperatures (Adamo et al., 2012). As study species we chose the damselfly *Ischnura elegans* and tested animals originating from different latitudes, hence with a different evolutionary history of heat extremes. Damselflies are especially sensitive to global warming (Hassall and Thompson, 2008) and given their aquatic larval stage and terrestrial adult stage, stressor effects after metamorphosis may couple

aquatic and terrestrial ecosystems (Stoks and Cordoba-Aguilar, 2012). We quantified the following fitness-related traits in damselflies (Stoks and Cordoba-Aguilar, 2012): survival, emergence success and development time in the larval stage and flying ability, body mass, water content, fat content, levels of the stress protein Hsp70 and phenoloxidase activity in the adult stage. As model pesticide we chose chlorpyrifos, an organophosphate insecticide that functions as an inhibitor of acetylcholinesterase (Stenersen, 2004). Chlorpyrifos is one of the most used pesticides worldwide (Eaton et al., 2008) and is often found in ponds where it negatively affects non-target organisms, including aquatic insects (Rubach et al., 2012).

1. Methods

1.1 Collecting and housing

We studied populations of the damselfly *Ischnura elegans* from two latitudes representing the low-latitude (southern France) and high-latitude (southern Sweden) parts of the range in Europe (Gosden et al., 2011). At each latitude three randomly chosen populations were sampled, namely Saint-Martin de Crau (+43°37'57.88"N, +4°46'55.18"E), Laune des Irudes (+43°30'34.80"N, +4°48'03.83"E) and Bassin de Réaltor (+43°28'11.16"N, +5°19'44.16"E) for France and Kalmar Dämme (+56°40'9.84"N, 16°17'48.48"E), Svino (+56°41'4.32"N, +16°22'23.79"E) and Hougårdsdammarna (+57°15'1.78"N, +12°8'1.983"E) for Sweden. These populations were chosen in natural areas without agriculture and it is therefore unlikely that they were exposed to pesticides (Coors et al., 2009). Furthermore, any local adaptation to pesticides would be unlikely in coenagrionid damselflies given their high levels of gene flow (Johansson et al., 2013). At both latitudes temperatures above 30 °C have been documented, yet such hot days are much more frequent in southern France compared to southern Sweden. While in southern France (weather station Tour-du-Valat) the average number of days with air

temperatures equal to or above 30 °C equaled 34 days per year during the period 1992-2008, this average was only one day per year in southern Sweden (weather station Malmö).

Eggs of fifteen mated females per population were collected and transferred to the laboratory in Belgium. One week after hatching, larvae were placed individually in 200 ml cups. Throughout the experiment, larvae were reared in incubators at 20 °C and L:D 14:10 hours. Larvae were daily fed *ad libitum* with *Artemia* nauplii five days a week (average daily dose = 1347 nauplii, SE = 102, $n = 15$ daily doses, each dose collected on a different day).

1.2 Pesticide concentration

Based on a previous experiment, we used pulses of 2.0 µg/l chlorpyrifos, since this causes a growth reduction and only limited mortality in *I. elegans* damselfly larvae (Dinh Van et al., in prep.). This concentration may be very high in general, but not in agricultural water bodies preferred by the study species (Dijkstra 2006). Here peak concentrations often exceed 100 µg/l due to runoff (e.g. Moore et al., 2002; Mazanti et al. 2003; Bernabo et al., 2011). We prepared the chlorpyrifos solution starting from a stock solution with a concentration of 20 µg/ml chlorpyrifos (kept in the dark at 4°C). This stock solution was a 50 times dilution of another stock solution containing 1 mg/ml chlorpyrifos dissolved in ethanol. The chlorpyrifos concentration of the stock solution at the start and at the end (3 months later) of the experiment was 1.000 mg/ml and 0.975 mg/ml, respectively. Samples were analyzed by the independent research laboratory Lovap NV (Geel, Belgium) using gas chromatography in combination with mass spectrometry. The initial chlorpyrifos concentration in the experimental vials was 2.020 µg/l and after three days (just before renewal of the medium) the concentration was lowered to 0.884 µg/l, indicating that although the chlorpyrifos concentration fluctuated in time, the larvae were continuously exposed to the pesticide.

We used aerated dechlorinated tap water in the control treatment instead of a solvent control since the amount of ethanol was only 2 µl/l exposure medium. A pilot experiment

showed that there was no difference in survival, growth and development time of the study species at this ethanol concentration (Janssens, unpublished results). Moreover, the lowest NOEC reported for aquatic invertebrates is >10,000 times higher than the ethanol concentrations used in the pesticide treatment (UNEP, 2004).

1.3 Experimental setup

To test for effects of larval pesticide exposure, larval food stress and adult heat exposure on survival, life history and physiological variables in *I. elegans* damselflies and compare those between two different latitudes, we set up a full factorial experiment with two latitudes (France versus Sweden, with three replicated populations per latitude) \times two food regimes (high food level versus low food level) \times three pesticide conditions (control, 16 days exposure, complete final instar exposure) \times two adult heat conditions (20 °C versus 30 °C) giving a total of 24 treatment combinations. We set up 35 larvae per treatment combination (total of 840 larvae). Due to differential mortality, sample sizes per response variable differ among treatment combinations and are shown in the figures.

The day after the larvae molted into the final instar (= F₀ stage), they entered the experiment. We used two different food levels: animals were fed *Artemia* nauplii twice a day six days a week (high food) or three days a week (low food); the individual doses per feeding were as before the exposure period (1347 nauplii \pm 102). We used two exposure durations to the pesticide: exposure during 16 days of the final instar (the shortest duration of the final instar) and exposure during the complete final instar. The fixed 16-d period guarantees an equal exposure duration of all animals across treatments and therefore allows testing for differences in susceptibility or toxicity (the effect of a chemical at a controlled exposure duration) among treatment combinations (Rohr et al., 2011). The treatment with exposure during the complete final instar reflects the scenario in nature where animals of different latitudes and at different food regimes have a different duration of the larval stage, which may

generate different exposure times to contaminants. Larvae were exposed in cups containing 100 ml of medium, which was renewed three times a week (static renewal experiment). The chronic exposure design reflects a realistic situation of runoff exposure in water bodies close to agricultural land where animals are exposed to several pesticide pulses per week during the growing season of the crops (Van Drooge et al., 2001).

The day after the adults emerged, half of them was exposed to heat (30 °C, 60-70 % humidity) for 24 h, the other half was kept at control conditions (20 °C, 60-70 % humidity) for 24h. The frequency with daily maximum temperatures exceeding 30 °C, is often used as an index to investigate changes in extreme temperature events (e.g. Beniston et al., 2007). We chose for a short pulse instead of a gradual change and longer exposure, since animals can physiologically acclimate to the latter (Kaur et al., 2011).

1.4 Response variables

We checked animals daily for survival and adult emergence. Animals were considered as survived when they were alive at emergence and emerged successfully (i.e. fully expanded their wings). Animals with not-fully expanded wings are unlikely to reach sexual maturity. Development time was calculated as the number of days between molting into the final instar and adult emergence. Two days after emergence (directly after the 24 h exposure at 20 °C or at 30 °C), each adult was weighed to the nearest 0.01 mg (electronic balance, Mettler Toledo® AB135-S), and the sex was determined. Additionally, we scored the flying ability (0: not able to fly, 1: able to fly) of each adult that emerged successfully in a small insectary (30 cm x 30 cm x 30 cm) by stimulating it by gently tapping on the abdomen. Afterwards, each adult was stored at -80 °C for physiological analyses.

To quantify the water content, an important trait in the terrestrial stage of arthropods to withstand arid conditions (Edney, 1977), and the lipid content, an important fitness-related trait in damselflies (Stoks and Cordoba-Aguilar, 2012), we followed the protocol of Swillen et

al. (2009). The thoraxes were placed individually in Eppendorf tubes and weighed to the nearest 0.1 µg using a microbalance (Thermo Cahn C-35) before and after being dried for 48 h at 60 °C. The water content was calculated as the difference between the wet mass and the dry mass and expressed as a percentage of the total wet mass. Fat was extracted by adding 1 ml dichloromethane (99%) to the Eppendorf tubes, which were shaken for 24 h on an automatic shaker. Thereafter, the thoraxes were dried for another 48 h at 60 °C and weighed. Fat content was quantified as the difference between the dry mass before and after fat extraction and expressed as a percentage of the total dry mass.

To quantify the levels of the stress protein Hsp70, which protects animals against heat stress (Sörensen et al., 2003), we used the western blot assay described in Slos and Stoks (2008). Shortly, we first homogenized the heads of the larvae in 80 µl phosphate buffer (PBS, 100 mM, pH 7.4) and measured the protein content of the samples using the Bradford method. Afterwards we diluted 30 µl of the head supernatant with 30 µl of Laemmli sample buffer and from this mixture a sample corresponding with 10 µg of protein was used for the analysis. The proteins were separated using SDS-polyacrylamide gel electrophoresis (PAGE). Afterwards, we used two antibodies to detect Hsp70, a monoclonal primary antibody (dilution 1:1500, anti-Hsp70 SPC-103D, Gentaur Europe, Kampenhout, Belgium) and an alkaline phosphatase-conjugated secondary antibody (dilution 1:1000, D0486, DakoCytomation, Glostrup, Denmark). We scanned the blotting membrane using the HP Scanjet 8270 and quantified the optical density of the stress protein bands using Image Pro Plus. Levels of Hsp70 were expressed as mean optical density (mOD) per µg protein. To correct for variation among blots, we ran on every blot a control sample of 1 µl Hela Cell Lysate (Heat shocked, Stressgen). Hsp70 levels were only quantified on a random subset of 12 larvae per treatment combination (total of 288 larvae).

We quantified the activity of phenoloxidase, a key component of an insect's immune system (Gonzalez-Santovo and Cordoba-Aguilar, 2012), based on the protocol of Stoks et al. (2006a). In a first step, we extracted the hemolymph by perfusing the thorax of the animals with 300 μ l phosphate buffer (PBS, 100 mM, pH 7.4). A 384 microtiter plate was filled with 40 μ l hemolymph and 5 μ l of chymotrypsin (5 mg/ml mili-Q water) and the mixture was incubated for 5 minutes at room temperature. This way all of the present pro-enzyme proPO was converted into PO. In a final step, we added 20 μ l L-DOPA substrate (10 mM in PBS). We measured absorbance at 490 nm during 60 minutes every 30 seconds at 30 °C. PO activity was determined as the slope of the linear part (between 1600 s and 2400 s) of the reaction curve. Measurements were run in duplicate per larva and the means were used for statistical analysis. Additionally, we measured protein content in the hemolymph sample using the Bradford method.

1.5 Statistical analyses

To test for effects of latitude of origin, larval pesticide exposure, larval food level, the adult heat treatment and their interactions on the binary response variables total survival and ability to fly we used separate ANOVAs with binomially distributed errors and the logit link function with the Glimmix procedure in SAS v9.3. To test for effects of latitude of origin, larval pesticide exposure, larval food level, the adult heat treatment and their interactions on the continuous variables dry mass, development time, water content and fat content we ran separate ANOVAs using the mixed procedure of SAS v9.3. When analyzing the PO activity and Hsp70 levels we used an ANCOVA with as covariates the protein content and body mass, and the optical density of the Hela band, respectively. We simplified the models by removing non-significant interactions and covariates. Population nested in latitude was included as a random factor. We also included sex in the models, but since it is no focal variable of interest and to keep the manuscript focused, we will only present its results in appendix 2. When a test

indicated a significant pesticide effect, we performed contrasts to investigate which of the three pesticide conditions differed from each other. The full models and the associated parameter estimates are presented in appendices 1 and 2 respectively. Means and standard errors per treatment combination for each response variable are given in appendix 3.

2. Results

2.1 Survival and flying ability

Survival was lower in Swedish (52 %) than in French animals (91 %) ($F_{1,4} = 80.76, p < 0.001$). No adults died during heat exposure, but afterwards the flying ability was lower ($F_{1,551} = 23.38, p < 0.001$; 20°C: 90% able to fly, 30°C: 67% able to fly); this pattern did not differ between French and Swedish animals (Latitude x Temperature: $F_{1,554} = 0.35, p = 0.56$). Food level, the pesticide nor interactions between the different stressors affected survival or flying ability.

2.2 Development time and mass at emergence

Development time (duration final instar) was longer in Swedish animals ($F_{1,607} = 2311.54, p < 0.0001$) and in animals reared at the low food level ($F_{1,607} = 20.67, p < 0.0001$). Swedish larvae exposed to the pesticide during the complete final instar had a longer development time at the high food level and a shorter development time at the low food level compared to the control larvae and the larvae exposed to the pesticide for 16 days. This was not the case in French animals (Latitude x Food x Pesticide; $F_{2,607} = 4.13, p = 0.017$) (Fig. 1A-B).

French adults had a lower dry mass than Swedish adults ($F_{1,5,85} = 13.69, p = 0.011$) and adults reared at the low larval food level were lighter than those reared at the high larval food level ($F_{1,441} = 113.31, p < 0.0001$). This reduction in dry mass at the low food level was more pronounced in French adults (Latitude x Food: $F_{1,441} = 17.53, p < 0.0001$) (Fig. 1C-D). Adult dry mass was lower when larvae had been exposed to chlorpyrifos ($F_{2,441} = 2.66, p =$

0.071; contrast analyses: $p_{\text{control-16days}} = 0.039$, $p_{\text{control-complete F0}} = 0.028$). There was no effect of the duration of the exposure period to chlorpyrifos on the dry mass (contrast analysis: $p_{16 \text{ days-complete F0}} = 0.88$). Dry mass tended to be lower in adults which were exposed to the heat treatment ($F_{1,441} = 3.71$, $p = 0.055$). This heat effect did not depend on latitude, food level or chlorpyrifos exposure (all $p < 0.34$) (Fig. 1C-D).

2.3 Physiology

Water content was higher in French adults than in Swedish adults ($F_{1,6,28} = 14.73$, $p = 0.0079$), and higher in French adults reared as larvae at the low food level than at the high food level (Latitude x Food: $F_{1,439} = 9.89$, $p = 0.0018$) (Fig. 2A-B). Animals that were exposed to chlorpyrifos during the complete final instar had the lowest water content ($F_{2,439} = 2.87$, $p = 0.058$; contrast analyses: $p_{\text{control-complete F0}} = 0.047$; $p_{16 \text{ days-complete F0}} = 0.039$). There was no difference in water content between the control animals and animals exposed to the pesticide for 16 days ($p_{\text{control-16days}} = 0.91$) (Fig. 2A-B). Water content was lower in adults which were exposed to heat compared with the control animals ($F_{1,440} = 34.85$, $p < 0.0001$).

Fat content was lower in French adults than in Swedish adults ($F_{1,5,88} = 158.42$, $p < 0.0001$). Animals reared at the low food level had a lower fat content ($F_{1,424} = 385.26$, $p < 0.0001$). Exposure to chlorpyrifos resulted in a lower fat content ($F_{2,425} = 178.6$, $p < 0.0001$), regardless of the exposure duration ($p_{16 \text{ days-complete F0}} = 0.90$). The pesticide-induced reduction of the fat content was more pronounced at the high food level (Pesticide x Food: $F_{2,424} = 32.18$, $p < 0.0001$) (Fig. 2C-D). Exposure to heat resulted in a lower fat content ($F_{1,425} = 157.41$, $p < 0.0001$), with a smaller reduction in adults that were exposed as larvae to the pesticide (Pesticide x Temperature: $F_{2,425} = 11.47$, $p < 0.0001$) (Fig. 2C-D).

Hsp70 levels were higher in Swedish adults ($F_{1,290} = 234.25$, $p < 0.001$) and lower in adults reared as larvae at the low food level ($F_{1,290} = 114.41$, $p < 0.001$). This decrease in Hsp70 levels at the low food level was more pronounced in the Swedish adults (Latitude x

Food: $F_{1, 290} = 4.05$, $p = 0.045$) (Fig. 3A-B). Exposure to the pesticide resulted in higher Hsp70 levels ($F_{2, 290} = 21.34$, $p < 0.001$), especially in animals reared at the high food level (Food \times Pesticide: $F_{2, 290} = 4.78$, $p = 0.0091$) (Fig. 3A-B). There was no effect of the exposure duration (contrast analysis: $p_{16 \text{ days-complete FO}} = 0.85$). Hsp70 levels were higher in adults which were exposed to heat ($F_{1, 290} = 48.16$, $p < 0.001$). This heat-induced increase in Hsp70 levels was more pronounced in Swedish adults (Latitude \times Temperature: $F_{1, 290} = 12.18$, $p < 0.001$), in animals reared at the high food level (Food \times Temperature: $F_{1, 290} = 8.19$, $p = 0.0045$), and in animals reared in the absence of the pesticide (Pesticide \times Temperature: $F_{2, 290} = 3.09$, $p = 0.047$) (Fig. 3A-B).

PO activity was higher in Swedish adults ($F_{1, 6.52} = 107.51$, $p < 0.0001$) and lower in adults reared as larvae at the low food level ($F_{1, 443} = 75.49$, $p < 0.0001$). Heat exposure caused a decrease in PO activity, but only in Swedish animals (Latitude \times Temperature: $F_{1, 443} = 9.95$, $p = 0.0017$) and in animals reared at the high food level (Food \times Temperature: $F_{1, 441} = 6.87$, $p = 0.0091$). Chlorpyrifos exposure resulted in a lower PO activity ($F_{2, 442} = 151.34$, $p < 0.0001$), while there was no effect of the exposure duration ($p_{16 \text{ days-complete FO}} = 0.43$). The pesticide-induced decrease in PO activity was the strongest in Swedish animals reared at the low food level (Food \times Pesticide \times Latitude: $F_{2, 441} = 6.01$, $p = 0.0027$) (Fig. 3C-D). The pesticide-induced decrease in PO activity was most pronounced in animals reared at the low food level and exposed to the heat (reduction of 53 %) and the least pronounced in animals reared at the high food level without heat exposure (reduction of 32 %) (Food \times Pesticide \times Temperature: $F_{2, 442} = 3.96$, $p = 0.020$, Figure 3C-D).

3. Discussion

Exposure to the ecologically relevant chlorpyrifos concentration in the larval stage of *I. elegans* affected nearly all sublethal traits scored after metamorphosis in the adult stage. As

shown in other studies, pesticide-exposed animals tended to be smaller at emergence (e.g. Janssens and Stoks, 2013a) and emerged as adults with lower energy reserves (e.g. Janssens and Stoks, 2013b) and lower immune function (e.g. Galloway and Handy, 2003). This can be explained by the investment in costly defence mechanisms in the presence of chlorpyrifos, including the here observed increase of Hsp70 (e.g. Janssens and Stoks, 2013b).

For most affected traits, the delayed effects of larval pesticide exposure on adult traits did not depend on the exposure duration. This suggests that longer exposure to the pesticide did not increase toxicity and that the animals were not able to recover (completely) from previous pesticide exposure, despite long durations of the larval post-exposure period in the 16-day exposure treatment (for example, the Swedish larvae that were exposed to the pesticide for 16 days had on average 56 days to recover before metamorphosis). Limited recovery at the individual level after pesticide exposure has been observed before (e.g. Rohr and Palmer, 2005) and is important when evaluating the effects of pesticides. This may not be surprising since the chosen chlorpyrifos concentration was already high compared to the $LC_{50_{48h}}$ for damselflies (5.28 $\mu\text{g/l}$, Brazner et al., 1988).

Only two traits were only affected when animals were exposed to the pesticide throughout the entire larval period: development time (see 4.2) and adult water content. We hypothesize the increased water content under long pesticide exposure to be a side-effect of an adaptive defense mechanism against pesticide uptake (Zhang et al., 2008), more specifically the reduction of the permeability of the cuticula by the incorporation of glycine-rich proteins (GRP) (Ahmad et al., 2006). The increase of GRP levels is energetically costly, what might explain the absence of a pesticide effect on the water content in animals that were only exposed for 16 days. The latter animals probably also decreased the permeability of the cuticula, but stopped GRP incorporation when the pesticide exposure ended.

3.1 Larval food stress and its interactions with pesticide stress

Animals reared at the low food level showed the typical slower development and reduction in mass at emergence (Nylin and Gotthard, 1998; De Block and Stoks, 2005), and invested less in costly traits such as fat content (Rolff et al., 2004; Stoks et al., 2006b; De Block et al., 2008), immune function (Rolff et al., 2004; De Block and Stoks, 2008; Karl et al., 2011) and the stress protein Hsp70 (e.g. Muturi et al., 2011).

Food level and pesticide exposure interacted for fat content. The pesticide-induced reduction of fat was more pronounced at high food; probably because high-food larvae can afford a stronger decrease in fat and invested more in the costly Hsp70 defense.

3.2 Latitude-specific responses to pesticide stress and food stress

We found strong phenotypic differentiation between the French and Swedish animals reflecting life history differences associated with latitudinal shifts in voltinism. The more favorable thermal conditions allow low-latitude populations to produce more than one generation per year, giving each generation, however, limited time to complete their life cycle thereby driving the evolution of a faster life history at low latitudes (e.g. Ragland and Kingsolver, 2007; for the study species: Shama et al., 2011; Stoks et al., 2012). This is here reflected in the shorter development times of the French larvae. A fast life history is, however, costly (Stoks et al., 2006b; Dmitriew, 2011) and is here traded off against a lower mass at emergence and a lower investment in Hsp70 (see also Stoks and De Block, 2011). We extended this fast-life phenotypic syndrome (*sensu* Réale et al., 2010) by also documenting lower investments in energy storage (fat content) and in immune function (PO activity). The higher investment in immune function in the Swedish animals in the absence of pathogens may explain their higher mortality (see Tucker and Stevens, 2003 for an example in beetles). The higher water content in French adults likely is an adaptation to the warm, dry conditions at low latitudes and is in line with the observation that insects adapted to hot and dry environments have a higher water content (e.g. Gibbs et al., 1997). A higher water content

may be achieved by a reduction of the permeability of the cuticula to prevent water loss by the incorporation of glycine-rich proteins (Zhang et al., 2008). Alternatively, it is possible that the French animals actively increased their water uptake to prevent emerging at a too small size, since this will have important negative fitness consequences (e.g. lower fecundity) (Stoks & Cordoba-Aguilar, 2012).

Despite strong latitudinal differences in nearly every variable in the unstressed control situation, French and Swedish animals responded quite similar to the pesticide stress. Yet, for two pesticide-affected variables, development time and PO activity, the French animals were less sensitive. When exposed to the pesticide during the complete final instar, French larvae did not change their development time, while Swedish larvae did so in a food-dependent way. At high food, Swedish larvae delayed emergence probably because under optimal food conditions this allowed them to compensate for the higher energy demands for detoxification (Sibly and Calow, 1989; including the here observed higher Hsp70 levels). Yet, at low food this was probably no option and they instead shortened the exposure period. Such adaptive shortening of the larval stage to escape pesticide exposure has been observed before (e.g. Distel and Boone, 2010; Rohr et al., 2011; Janssens and Stoks, 2013a). The absence of this life history escape response in French larvae probably reflects a lower limit to their, already very short, final instar (see also Stoks and De Block, 2011). Somewhat similar, the pesticide-induced immunosuppression was stronger in Swedish adults reared as larvae at low food. As we will argue for heat exposure, the high PO levels in Swedish animals may have relaxed the need to buffer the pesticide-induced immunosuppression at low food. The smaller response in French animals can be explained by their lower baseline PO levels that probably were close to the admissible level to survive immune challenges (see also Campero et al., 2008).

In line with the observation that faster growing animals invest less in energy storage and are more sensitive to food shortage (e.g. Stoks et al., 2006b), the food stress-induced

reduction in mass was stronger in French animals; this despite French adults already being lighter than the Swedish adults in the unstressed condition. In contrast, however, the food stress-induced reduction in Hsp70 was less strong in French animals, suggesting they tried to keep Hsp70 levels above a certain critical threshold, potentially to cope with future thermal stress. Only French animals increased their water content when reared at the low food level (for an example on collembolan, see Lavy et al., 1997). Adult size is fixed at emergence in insects and important for adult fitness (Stoks and Cordoba-Aguilar, 2012). We hypothesize that French animals reared at low food increased water uptake to compensate any size reductions due to the loss of mass and fat reserves.

3.3 Adult heat exposure and its effect on carry-over effects of larval pesticide stress

Heat exposure had strong negative effects on nearly every trait, suggesting it will negatively impact fitness. Besides the direct effect of a lowered water content (e.g. Atmowidjojo et al., 1997), heat-exposed animals suffered reductions in mass (e.g. Franke and Fischer, 2013), fat content and flying ability (e.g. Wang et al., 2009). This can be explained by the costly defense mechanisms against the heat treatment, including the here observed increase of Hsp70 levels (Sörensen et al., 2003). Additionally, heat exposure likely increased metabolic rate, thereby generating higher energy demands (Clarke and Fraser, 2004). Noteworthy, while it has been recently demonstrated that effects of an extreme temperature may be stronger at low food level (Adamo et al., 2012), this was not supported in our study.

In line with the much higher frequencies of extreme temperatures at their latitude, we found some indication that French adults could better deal with the extreme temperature. French adults responded with a weaker increase in Hsp70 in response to the heat treatment. We interpret this as the French larvae being more adapted to extreme temperatures, making it less necessary to rely on the energetically costly increase of Hsp70 (see e.g. Tomanek & Zuzow, 2010 for an example in mussels). Furthermore, only Swedish adults reduced PO

levels after the heat exposure. Yet, the latter effect may not necessarily reflect a lower ability to deal with heat stress, but rather the fact that Swedish adults can better afford a decrease in PO (given their higher levels in the unstressed controls). This may also explain why the heat-induced reduction in PO was only observed at high food. This adds to the few other studies showing heat-induced reductions in PO activity (Karl et al., 2011; Seppälä and Jokela, 2011), thereby indicating that extreme temperatures, which are predicted to increase in frequency under global warming (IPCC, 2012), may make animals more susceptible to pathogens.

A key finding was that adult heat exposure modulated the carry-over costs of the larval pesticide exposure, and this in opposite ways for fat content and PO activity. The pesticide-induced reduction in adult fat content was less strong in adults exposed to the heat treatment. This again can be interpreted as adults in treatment groups with higher values of a fitness-related trait (here fat content in adults not exposed to the heat treatment) can afford stronger reductions in the trait levels without going below a critical threshold value. In contrast, the pesticide-induced reduction in PO activity (at low food level) was stronger in heat-exposed adults. This can be explained as a physiological cost of the stress exposure: adults that were exposed to the pesticide during the larval stage, had probably a lower energy content which was invested in defense against the stressor that was present at that moment (i.e. heat) instead of investing it in long-term defense (i.e. immune response).

Larval pesticide exposure and larval food stress also reduced the defense response to the adult heat stress in terms of increased Hsp70 levels. This is likely the result of their lower energy content due to a lower energy income and/or need to invest in detoxification. This may also have contributed to the pattern of lowest immune function (PO activity) in heat-exposed adults that were exposed as larvae to a combination of pesticide and food stress.

3.4 Conclusions and implications for ecological risk assessment under global warming

A key challenge for ecotoxicologists is to assess the impact of pesticides under global warming (Noyes et al. 2009, Moe et al. 2013). By studying how carry-over effects of larval pesticide exposure across metamorphosis are modulated by a subsequent adult heat exposure in populations along opposite ends of a natural thermal gradient, our study highlighted two novel key findings relevant for ecological risk assessment under global warming (Landis et al., 2013). Firstly, larval pesticide stress and adult heat stress interacted across metamorphosis making the delayed effects of larval pesticide exposure dependent upon subsequent adult heat exposure for two fitness-related traits (fat and PO at low food). Related to this, animals subsequently exposed to both stressors across metamorphosis suffered the lowest values of these traits, making it relevant to consider these combined effects across metamorphosis when predicting the fate of populations in polluted sites under global warming.

Secondly, in line with strong differentiation in the unstressed control situation, French and Swedish animals responded differently to the imposed stressors. While the fast growing French animals were more sensitive to larval food stress (for body mass and water content), they were less responsive to the pesticide stress (for development time and PO activity) and heat exposure (for PO activity and Hsp70 levels) compared to the Swedish animals. This indicated that French suffered more under lower energy acquisition, while under an energy allocation trade-off they better succeeded in maintaining minimum investment in immune function and heat shock defense. These results highlight that intraspecific life history evolution along natural thermal gradients may shape sensitivity to a pesticide and therefore should be included in ecological risk assessment under global warming. More general our results support the recent plea to include evolutionary developed phenotypic attributes when considering the effects of contaminants in aquatic systems (Kamo et al., 2011; Segner, 2011).

Acknowledgements

We thank two anonymous reviewers for their constructive comments that improved the manuscript. We thank Philippe Lambret and Viktor Nilsson-Örtman for collecting eggs and Gregory Malfait for assisting during the experiment. LJ is a PhD fellow of the IWT, Flanders. A collection and rearing permit for damselflies was obtained from ANB-Flanders. Financial support was provided by research projects of FWO-Flanders and the KU Leuven Centre of Excellence grant PF/2010/07. We declare no conflict of interest.

Supplementary data

Appendix 1: Parameter estimates and their errors of the AN(C)OVAs testing for effects of latitude, food level, pesticide exposure and heat exposure on fitness-related traits in damselflies.

Appendix 2: Results of the full model AN(C)OVAs testing for effects of latitude, food level, pesticide exposure and heat exposure on fitness-related traits in damselflies.

Appendix 3: Means and their standard errors of the combined impact of latitude, food level, pesticide exposure and heat exposure on the continuous fitness related traits development time, dry mass, water content, fat content, Hsp70 levels and PO activity.

References

- Adamo, S.A., Baker, J.L., Lovett, M.M.E., Wilson, G., 2012. Climate change and temperate zone insects: the tyranny of thermodynamics meets the world of limited resources. *Environmental Entomology* 41, 1644-1652.
- Ahmad, M., Denholm, I., Bromilow, R.H., 2006. Delayed cuticular penetration and enhanced metabolism of deltamethrin in pyrethroid-resistant strains of *Helicoverpa armigera* from China and Pakistan. *Pest Management Science* 62, 805-810.

496 Atmowidjojo, A.H., Wheeler, D.E., Erickson, E.H., Cohen, A.C., 1997. Temperature
 497 tolerance and water balance in feral and domestic honey bees, *Apis mellifera* L.
 498 Comparative Biochemistry and Physiology A 118, 1399-1403.

499 Beniston, M., Stephenson, D.B., Christensen, O.B., Ferro, C.A.T., Frei, C., Goyette, S.,
 500 Halsnaes, K., Holt, T., Jylhä, K., Koffi, B., Palutikoff, J., Schöll, R., Semmler, T., Woth,
 501 K., 2007. Future extreme events in European climate: an exploration of Regional Climate
 502 Model projections. Climate Change 81, 71-95.

503 Bernabo, I., Sperone, E., Tripepi, S., Brunelli, E., 2011. Toxicity of chlorpyrifos to larval
 504 *Rana dalmatina*: acute and chronic effects on survival, development, growth and gill
 505 apparatus. Archives of Environmental Contamination and Toxicology 61, 704-718.

506 Bronmark, C., Hansson, L.A., 2002. Environmental issues in lakes and ponds: current state
 507 and perspectives. Environmental Conservation 29, 290-307.

508 Boone, M.D., Hammond, S.A., Veldhoen, N., Youngquist, M., Helbing, C.C., 2013. Specific
 509 time of exposure during tadpole development influences biological effects of the
 510 insecticide carbaryl in green frogs (*Lithobates clamitans*). Aquatic Toxicology 130, 139-
 511 148.

512 Brazner, J.C., Lozaio, S.J., Knuth, M.L., Bertelsen, S.L., Heinis, L.J., Jensen, D.A., Kline,
 513 E.R., O'Halloran, S.L., Sargent, K.W., Tanner, O.K., 1988. The effects of chlorpyrifos on a
 514 natural aquatic system: a research design for littoral enclosure studies and final research
 515 report. Final report. US Environmental Protection Agency. Duluth, MN.

516 Campero, M., De Block, M., Ollevier, F., Stoks, R., 2008. Correcting the short-term effect of
 517 food deprivation in a damselfly: mechanisms and costs. Journal of Animal Ecology 77, 66-
 518 73.

519 Clarke, A., Fraser, K.P.P., 2004. Why does metabolism scale with temperature? Functional
 520 Ecology 18, 243-251.

521 Clements, W.H., Hickey, C.W., Kidd, K.A., 2012. How do aquatic communities respond to
 522 contaminants? It depends on the ecological context. *Environmental Toxicology and*
 523 *Chemistry* 31, 1932-1940.

524 Coors, A., Vanoverbeke, J., De Bie, T., De Meester, L., 2009. Land use, genetic diversity and
 525 toxicant tolerance in natural populations of *Daphnia magna*. *Aquatic Toxicology* 95, 71-
 526 79.

527 De Block, M., Slos, S., Johansson, F., Stoks, R., 2008. Integrating life history and physiology
 528 to understand latitudinal size variation in a damselfly. *Ecography* 31, 115-123.

529 De Block, M., Stoks, R., 2005. Fitness effects from egg to reproduction: bridging the life
 530 history transition. *Ecology* 86, 185-197.

531 De Block, M., Stoks, R., 2008. Short-term larval food stress and associated compensatory
 532 growth reduce adult immune function in a damselfly. *Ecological Entomology* 33, 796-801.

533 Deutsch, C.A., Tewksbury, J.J., Huey, R.B., Sheldon, K.S., Ghalambor, C.K., Haak, D.C.,
 534 Martin, P.R., 2008. Impacts of climate warming on terrestrial ectotherms across latitude.
 535 *Proceedings of the National Academy of Sciences USA* 105, 6668-6672.

536 Dijkstra, K.D.B., 2006. Field guide to the dragonflies of Britain and Europe. British Wildlife
 537 Publishing, Dorset, UK.

538 Distel, C.A., Boone, M.D., 2010. Effects of aquatic exposure to the insecticide carbaryl are
 539 species-specific mediated by heterospecific competitors in anurans. *Functional Ecology* 24,
 540 1342-1352.

541 Dmitriew, C.M., 2011. The evolution of growth trajectories: what limits growth rate?
 542 *Biological Reviews* 86, 97-116.

543 Eaton, D.L., Daroff, R.B., Autrup, H., Bridges, J., Buffler, P., Costa, L.G., Coyle, J.,
 544 McKhann, G., Mobley, W.C., Nadel, L., Neubert, D., Schulte-Hermann, R., Spencer, P.S.,

545 2008. Review of the toxicology of chlorpyrifos with emphasis on human exposure and
 546 neurodevelopment. *Critical Reviews in Toxicology* 38, 1-125.
 547 Edney, E.B., 1977. Water balance in land arthropods. *Zoophysiology and Ecology*. Volume 9,
 548 Springer, London, UK.
 549 Franke, K., Fischer, K., 2013. Effects of inbreeding and temperature stress on life history and
 550 immune function in a butterfly. *Journal of Evolutionary Biology* 26, 517-528.
 551 Galloway, T., Handy, R., 2003. Immunotoxicity of organophosphorous pesticides.
 552 *Ecotoxicology* 12, 345-363.
 553 Gibbs, A.G., Chippindale, A.K., Rose, M.R., 1997. Physiological mechanisms of evolved
 554 desiccation resistance in *Drosophila melanogaster*. *Journal of Experimental Biology* 200,
 555 1821-1832.
 556 González-Santoyo, I., Córdoba-Aguilar, A., 2012. Phenoloxidase: a key component of the
 557 insect immune system. *Entomologia Experimentalis et Applicata* 142, 1-16.
 558 Gosden, T.P., Stoks, R., Svensson, E.I., 2011. Range limits, large-scale biogeographic
 559 variation and localized evolutionary dynamics in a polymorphic damselfly. *Biological*
 560 *Journal of the Linnean Society* 102, 775-785.
 561 Hassal, C., Thompson, D.J., 2008. The effects of environmental warming on Odonata: a
 562 review. *International Journal of Odonatology* 11, 131-153.
 563 Hoffmann, A.A., Anderson, A., Hallas, R., 2002. Opposing clines for high and low
 564 temperatures in *Drosophila melanogaster*. *Ecology Letters* 5, 614-618.
 565 Intergovernmental Panel on Climate Change, 2007. *Climate change 2007: the physical*
 566 *science basis*. Cambridge University Press, Cambridge, UK.
 567 Intergovernmental Panel on Climate Change, 2012. *Managing the risks of extreme events and*
 568 *disaster to advance climate change adaptation*. Cambridge University Press, Cambridge,
 569 UK.

570 Janssens, L., Stoks, R., 2013a. Fitness effects of chlorpyrifos in the damselfly *Enallagma*
571 *cyathigerum* strongly depend upon temperature, food level and the fitness-related variable.
572 PLoS One 8, e68107.

573 Janssens L., Stoks R., 2013b. Exposure to a widespread non-pathogenic bacterium magnifies
574 sublethal pesticide effects in the damselfly *Enallagma cyathigerum*: from the
575 suborganismal level to fitness-related traits. Environmental Pollution 177, 143-149.

576 Jentsch, A., Kreyling, J., Beierkuhnlein, C., 2007. A new generation of climate change
577 experiments: events, not trends. Frontiers in Ecology and the Environment 5, 365-374.

578 Johansson, H., Stoks, R., Nilsson-Ortman, V., Ingvarsson, P.K., Johansson, F., 2013. Large-
579 scale patterns in genetic variation, gene flow and differentiation in five species of
580 European Coenagrionid damselfly provide mixed support for the central-marginal
581 hypothesis. Ecography 36, 744-755.

582 Kamo, M., Hayashi, T.I., Akita, T., 2011. Potential effects of life-history evolution on
583 ecological risk assessment. Ecological Applications 21, 3191-3198.

584 Karl, I., Stoks, R., De Block, M., Janowitz, S.A., Fischer, K., 2011. Temperature extremes
585 and butterfly fitness: conflicting evidence from life history and immune function. Global
586 Change Biology 17, 676-687.

587 Kaur, M., Atif, F., Ansari, R.A., Ahmad, F., Raisuddin, S., 2011. The interactive effect of
588 elevated temperature on deltamethrin-induced biochemical stress responses in *Channa*
589 *punctata* Bloch. Chemico-Biological Interactions 193, 216-224.

590 Landis, W.G., Durda, J.L., Brooks, M.L., Chapman, P.M., Menzie, C.A., Stahl, R.G., Stauber,
591 J.L., 2013. Ecological risk assessment in the context of global climate change.
592 Environmental Toxicology and Chemistry 32, 79-92.

593 Lavy, D., Nedved, O., Verhoef, H.A., 1997. Effects of starvation on body composition and
 594 cold tolerance in the collembolan *Orchesella cincta* and the isopod *Porcellio scaber*.
 595 Journal of Insect Physiology 43, 973-978.

596 Mazanti, L., Rice, C., Bialek, K., Sparling, D., Stevenson, C., Johnson, W.E., Kangas, P.,
 597 Rheinstein, J., 2003. Aqueous-phase disappearance of atrazine, metolachlor and chlorpyrifos
 598 in laboratory aquaria and outdoor macrocosms. Archives of Environmental Contamination
 599 and Toxicology 44, 67-76.

600 Moe, S.J., De Schamphelaere, K., Clements, W.H., Sorensen, M.T., Van den Brink, P.J.,
 601 Liess, M., 2013. Combined and interactive effects of global climate change and toxicants
 602 on populations and communities. Environmental Toxicology and Chemistry 32, 49-61.

603 Moore, M.T., Schulz, R., Cooper, C.M., Smith Jr., S., Rodgers Jr., J.H., 2002. Mitigation of
 604 chlorpyrifos runoff using constructed wetlands. Chemosphere 46, 827-835.

605 Moran, N.A., 1994. Adaptation and constraint in the complex life-cycle of animals. Annual
 606 Review of Ecology and Systematics 25, 573-600.

607 Muturi, E.J., Kim, C.H., Alto, B.W., Berenbaum, M.R., Schuler, M.A., 2011. Larval
 608 environmental stress alters *Aedes aegypti* competence for Sindbis virus. Tropical Medicine
 609 & International Health 16, 955-964.

610 Noyes, P.D., McElwee, M.K., Miller, H.D., Clark, B.W., Van Tiem, L.A., Walcott, K.C.,
 611 Erwin, K.N., Levin, E.D., 2009. The toxicology of climate change: Environmental
 612 contaminants in a warming world. Environment International 35, 971-986.

613 Nylin, S., Gotthard, K., 1998. Plasticity in life-history traits. Annual Review of Entomology
 614 43, 63-83.

615 Orlowsky, B., Seneviratne, S.I., 2012. Global changes in extreme events: regional and
 616 seasonal dimension. Climate Change 110, 669-696.

617 Ragland, G.J., Kingsolver, J.G., 2008. Evolution of thermotolerance in seasonal
 618 environments: The effects of annual temperature variation and life-history timing in
 619 *Wyeomyia smithii*. *Evolution* 62, 1345-1357.

620 Réale, D., Garant, D., Humphries, M.M., Bergeron, P., Careau, V., Montiglio, P.O., 2010.
 621 Personality and the emergence of the pace-of-life syndrome concept at the population
 622 level. *Philosophical Transactions of the Royal Society B* 365, 4051-4063.

623 Rohr, J.R., Palmer, B.D., 2005. Aquatic herbicide exposure increases salamander desiccation
 624 risk eight months later in a terrestrial environment. *Environmental Toxicology and*
 625 *Chemistry* 24, 1253-1258.

626 Rohr, J.R., Sesterhenn, T.M., Stieha, C., 2011. Will climate change reduce the effects of a
 627 pesticide on amphibians?: partitioning effects of exposure and susceptibility to
 628 contaminants. *Global Change Biology* 17, 657-666.

629 Rolff, J., Van de Meutter, F., Stoks, R., 2004. Time constraints decouple age and size at
 630 maturity and physiological traits. *American Naturalist* 164, 559-565.

631 Rubach, M.N., Baird, D.J., Boerwinkel, M.C., Maund, S.J., Roessink, I., Van den Brink, P.J.,
 632 2012. Species traits as predictors for intrinsic sensitivity of aquatic invertebrates to the
 633 insecticide chlorpyrifos. *Ecotoxicology* 21, 2088-2101.

634 Segner, H., 2011. Moving beyond a descriptive aquatic toxicology: the value of biological
 635 process and trait information. *Aquatic Toxicology* 105, 50-55.

636 Seppälä, O., Jokela, J., 2011. Immune defence under extreme ambient temperature. *Biology*
 637 *Letters* 7, 119-122.

638 Shama, L.N.S., Campero-Paz, M., Wegner, K.M., De Block, M., Stoks, R., 2011. Latitudinal
 639 and voltinism compensation shape thermal reaction norms for growth rate. *Molecular*
 640 *Ecology* 20, 2929-2941.

641 Sibly, R.M., Calow, P., 1989. A life-cycle theory of response to stress. *Biological Journal of*
642 *the Linnean Society* 37, 101-116.

643 Slos, S., Stoks, R., 2008. Predation risk induces stress proteins and reduces antioxidant
644 defense. *Functional Ecology* 22, 637-642.

645 Sörensen, J.G., Kristensen, T.N., Loeschcke, V., 2003. The evolutionary and ecological role
646 of heat shock proteins. *Ecology Letters* 6, 1025-1037.

647 Stenersen, J., 2004. Chemical pesticides: mode of action and toxicology. CRC Press, Boca
648 Raton, Florida.

649 Stoks, R., Cordoba-Aguilar, A., 2012. Evolutionary ecology of Odonata: a complex life cycle
650 perspective. *Annual Review of Entomology* 57, 249-265.

651 Stoks, R., De Block, M., 2011. Rapid growth rate reduces cold resistance: evidence from
652 latitudinal variation in growth rate, cold resistance and stress proteins. *PLoS One* 6,
653 e16935.

654 Stoks, R., De Block, M., McPeck, M.A., 2006b. Physiological costs of compensatory growth
655 in a damselfly. *Ecology* 87, 1566-1574.

656 Stoks, R., De Block, M., Slos, S., Van Doorslaer, W., Rolff, J., 2006a. Time constraints
657 mediate predator-induced plasticity in immune function, condition and life history.
658 *Ecology* 87, 809-815.

659 Stoks, R., Swillen, I., De Block, M., 2012. Behaviour and physiology shape the growth
660 accelerations associated with predation risk, high temperatures and southern latitudes in
661 *Ischnura* damselfly larvae. *Journal of Animal Ecology* 81, 1034-1040.

662 Swillen, I., De Block, M., Stoks, R., 2009. Morphological and physiological sexual selection
663 targets in a territorial damselfly. *Ecological Entomology* 34, 677-683.

664 Tomanek, L., Zuzow, M.J., 2010. The proteomic response of the mussel congeners *Mytilus*
665 *galloprovincialis* and *M. trossulus* to acute heat stress: implications for thermal tolerance

limits and metabolic costs of thermal stress. *Journal of Experimental Biology* 213, 3559-3574.

Tucker, T.M., Stevens, L., 2003. Geographical variation and sexual dimorphism of phenoloxidase levels in Japanese beetles (*Popillia japonica*). *Proceedings of the Royal Society B* 270, S245-S247.

United Nations Environment Program (UNEP), 2004. Screening Information DataSet: initial assessment report ethanol cas N°: 64-17-5.

Van Drooge, H.L., Groenvelde, C.N., Schipper, H.J., 2001. Data on application frequency of pesticide for risk assessment purposes. *Annals of Occupational Hygiene* 45, S95-S101.

Wang, X.G., Johnson, M.W., Daane, K.M., Opp, S., 2009. Combined effects of heat stress and food supply on flight performance of olive fruit fly (Diptera: Tephritidae). *Entomological Society of America* 102, 727-734.

Woodward, G., Perkins, D.M., Brown, L.E., 2010. Climate change and freshwater ecosystems: impacts across multiple levels of organization. *Philosophical Transactions of the Royal Society B* 365, 2093-2106.

Zhang, J., Goyer, C., Pelletier, Y., 2008. Environmental stress induce the expression of putative glycine-rich insect cuticular protein genes in adult *Leptinotarsa decemlineata* (Say). *Insect Molecular Biology* 17, 209-216.

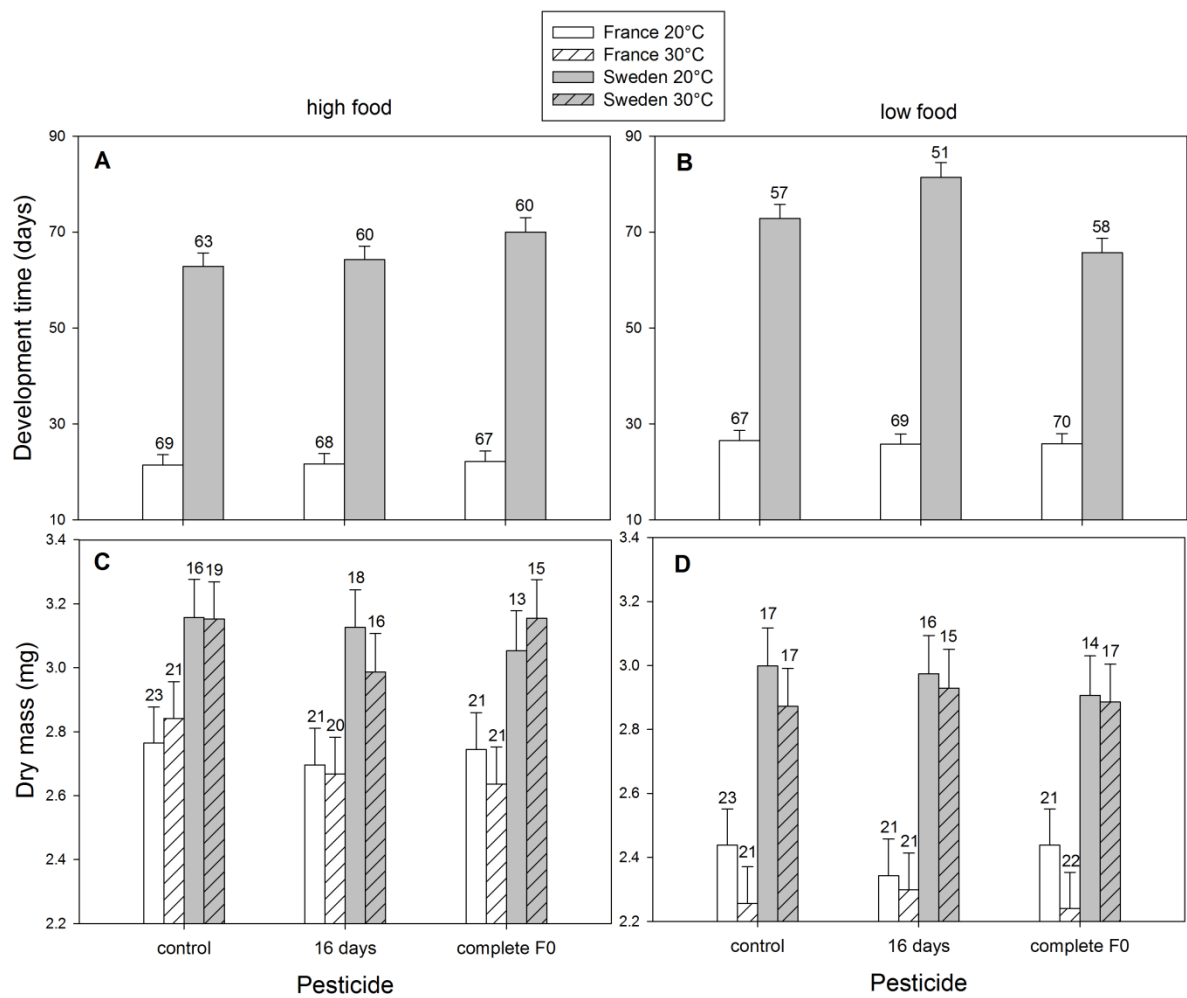
Figure legends

Fig. 1: Mean development time (A, B) and dry mass (C, D) of *I. elegans* damselflies as a function of latitude of origin, larval food stress, larval pesticide exposure and adult heat exposure. Given are least-squares means \pm 1 SE.

Fig. 2: Mean water content (A, B) and fat content (C, D) of *I. elegans* damselflies as a function of latitude of origin, larval food stress, larval pesticide exposure and adult heat exposure. Given are least-squares means \pm 1 SE.

Fig. 3: Mean Hsp70 levels (A, B) and phenoloxidase activity (C,D) of *I. elegans* damselflies as a function of latitude of origin, larval food stress, larval pesticide exposure and adult heat exposure. Given are least-squares means \pm 1 SE.

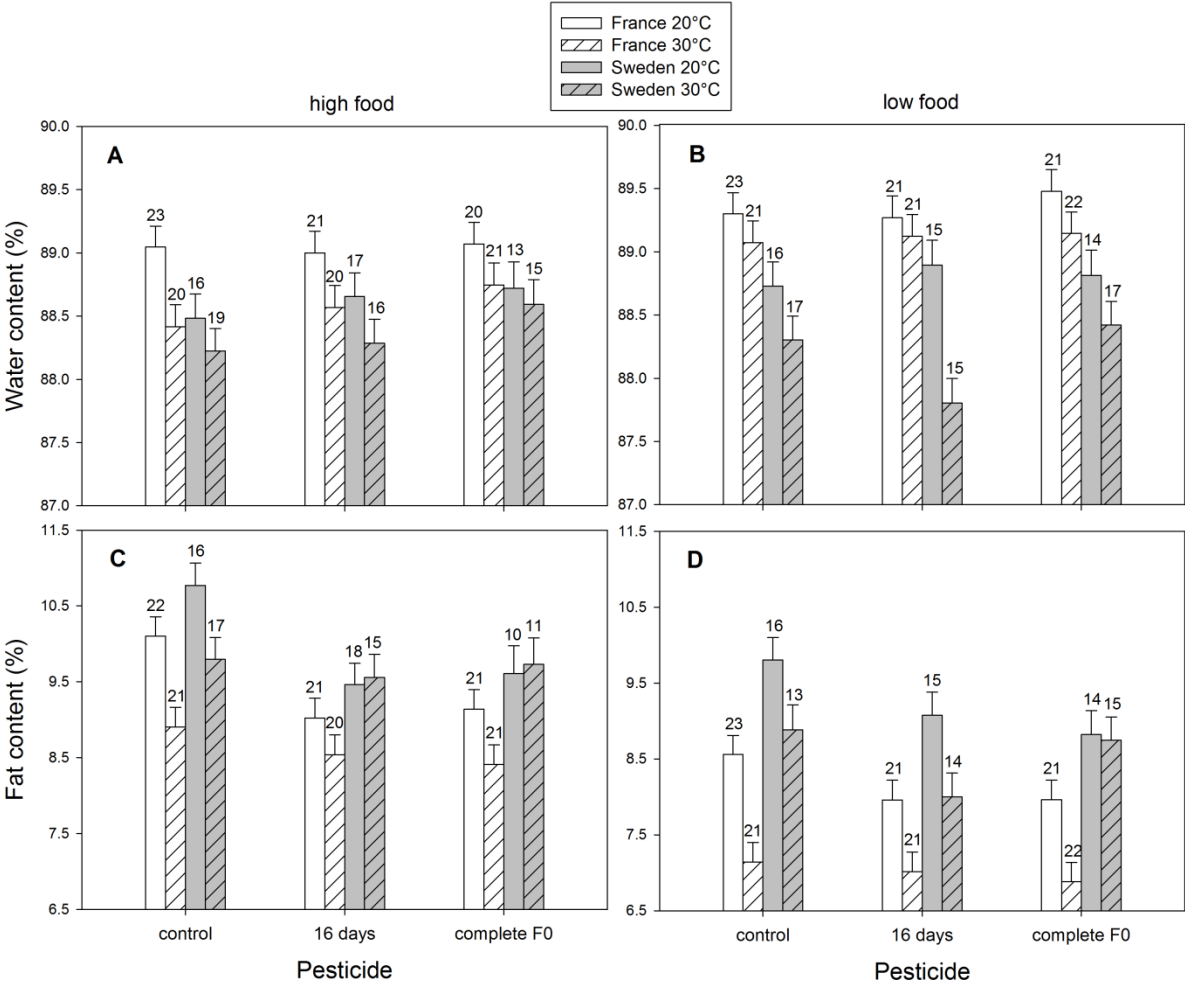
701 **Fig. 1**



702

703

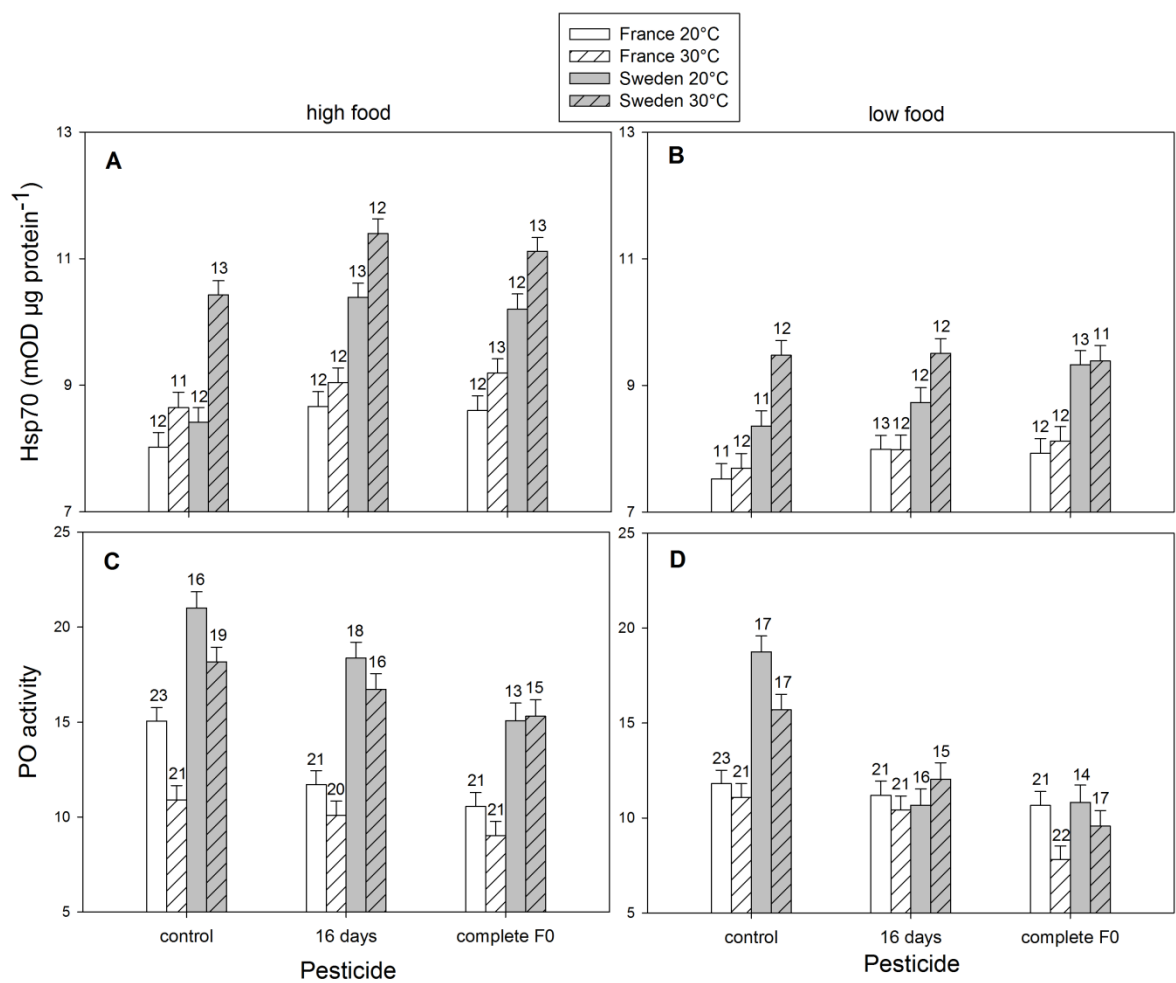
704 **Fig. 2**



705

706

Fig. 3



710 Appendix 1. Parameter estimates and their standard errors of the ANCOVAs reported in the manuscript.

711

Response variable	Effect	Sex	Latitude	Food	Pesticide	Heat	Estimate	Standard Error
Survial	Latitude (L)		France				17.434	0.6099
	Latitude (L)		Sweden				0	.
	Food (F)			high			-0.3105	0.4907
	Food (F)			low			0	.
	Pesticide (P)				control		0.3821	0.5148
	Pesticide (P)				16 days		-0.2495	0.4940
	Pesticide (P)				complete F0		0	.
	Heat (H)					20°C	-0.1715	0.4898
	Heat (H)					30°C	0	.
	L * F		France	high			19.373	12.260
	L * F		France	low			0	.
	L * F		Sweden	high			0	.
	L * F		Sweden	low			0	.
	L * P		France		control		0.08313	0.9287
	L * P		France		16 days		0.2683	0.8403
	L * P		France		complete F0		0	.
	L * P		Sweden		control		0	.
	L * P		Sweden		16 days		0	.
	L * P		Sweden		complete F0		0	.
	L * H		France			20°C	0.6718	0.9145
	L * H		France			30°C	0	.
	L * H		Sweden			20°C	0	.
	L * H		Sweden			30°C	0	.
	F * P			high	control		0.03914	0.7073
	F * P			high	16 days		0.4429	0.7014

F * P		high	complete F0		0	.
F * P		low	control		0	.
F * P		low	16 days		0	.
F * P		low	complete F0		0	.
F * H		high		20°C	-0.02337	0.6941
F * H		high		30°C	0	.
F * H		low		20°C	0	.
F * H		low		30°C	0	.
P * H			control	20°C	-0.3220	0.7103
P * H			control	30°C	0	.
P * H			16 days	20°C	0.3724	0.6930
P * H			16 days	30°C	0	.
P * H			complete F0	20°C	0	.
P * H			complete F0	30°C	0	.
L * F * P	France	high	control		-21.658	15.366
L * F * P	France	high	16 days		-11.808	15.863
L * F * P	France	high	complete F0		0	.
L * F * P	France	low	control		0	.
L * F * P	France	low	16 days		0	.
L * F * P	France	low	complete F0		0	.
L * F * P	Sweden	high	control		0	.
L * F * P	Sweden	high	16 days		0	.
L * F * P	Sweden	high	complete F0		0	.
L * F * P	Sweden	low	control		0	.
L * F * P	Sweden	low	16 days		0	.
L * F * P	Sweden	low	complete F0		0	.
L * F * H	France	high		20°C	-18.882	15.461
L * F * H	France	high		30°C	0	.
L * F * H	France	low		20°C	0	.
L * F * H	France	low		30°C	0	.

L * F * H	Sweden	high		20°C	0	.
L * F * H	Sweden	high		30°C	0	.
L * F * H	Sweden	low		20°C	0	.
L * F * H	Sweden	low		30°C	0	.
L * P * H	France		control	20°C	0.9832	15.803
L * P * H	France		control	30°C	0	.
L * P * H	France		16 days	20°C	-0.7060	12.609
L * P * H	France		16 days	30°C	0	.
L * P * H	France		complete F0	20°C	0	.
L * P * H	France		complete F0	30°C	0	.
L * P * H	Sweden		control	20°C	0	.
L * P * H	Sweden		control	30°C	0	.
L * P * H	Sweden		16 days	20°C	0	.
L * P * H	Sweden		16 days	30°C	0	.
L * P * H	Sweden		complete F0	20°C	0	.
L * P * H	Sweden		complete F0	30°C	0	.
F * P * H		high	control	20°C	0.6902	0.9942
F * P * H		high	control	30°C	0	.
F * P * H		high	16 days	20°C	0.6226	0.9966
F * P * H		high	16 days	30°C	0	.
F * P * H		high	complete F0	20°C	0	.
F * P * H		high	complete F0	30°C	0	.
F * P * H		low	control	20°C	0	.
F * P * H		low	control	30°C	0	.
F * P * H		low	16 days	20°C	0	.
F * P * H		low	16 days	30°C	0	.
F * P * H		low	complete F0	20°C	0	.
F * P * H		low	complete F0	30°C	0	.
L * F * P * H	France	high	control	20°C	0.6287	22.113
L * F * P * H	France	high	control	30°C	0	.

L * F * P * H	France	high	16 days	20°C	11.165	21.162
L * F * P * H	France	high	16 days	30°C	0	.
L * F * P * H	France	high	complete F0	20°C	0	.
L * F * P * H	France	high	complete F0	30°C	0	.
L * F * P * H	France	low	control	20°C	0	.
L * F * P * H	France	low	control	30°C	0	.
L * F * P * H	France	low	16 days	20°C	0	.
L * F * P * H	France	low	16 days	30°C	0	.
L * F * P * H	France	low	complete F0	20°C	0	.
L * F * P * H	France	low	complete F0	30°C	0	.
L * F * P * H	Sweden	high	control	20°C	0	.
L * F * P * H	Sweden	high	control	30°C	0	.
L * F * P * H	Sweden	high	16 days	20°C	0	.
L * F * P * H	Sweden	high	16 days	30°C	0	.
L * F * P * H	Sweden	high	complete F0	20°C	0	.
L * F * P * H	Sweden	high	complete F0	30°C	0	.
L * F * P * H	Sweden	low	control	20°C	0	.
L * F * P * H	Sweden	low	control	30°C	0	.
L * F * P * H	Sweden	low	16 days	20°C	0	.
L * F * P * H	Sweden	low	16 days	30°C	0	.
L * F * P * H	Sweden	low	complete F0	20°C	0	.
L * F * P * H	Sweden	low	complete F0	30°C	0	.

Flying ability	Sex	female			0.6018	0.2315
	Sex	male			0	.
	Latitude (L)		France		-2.1201	1.1015
	Latitude (L)		Sweden		0	.
	Food (F)			high	-0.1607	1.4662
	Food (F)			low	0	.
	Pesticide (P)			control	-2.5567	1.1413

Pesticide (P)			16 days	-3.0253	1.1703
Pesticide (P)			complete F0	0	.
Heat (H)			20°C	-0.8390	1.2841
Heat (H)			30°C	0	.
L * F	France	high		0.5544	.
L * F	France	low		0	.
L * F	Sweden	high		0	.
L * F	Sweden	low		0	.
L * P	France		control	2.6063	1.2623
L * P	France		16 days	2.7397	1.2801
L * P	France		complete F0	0	.
L * P	Sweden		control	0	.
L * P	Sweden		16 days	0	.
L * P	Sweden		complete F0	0	.
L * H	France		20°C	2.4270	1.4699
L * H	France		30°C	0	.
L * H	Sweden		20°C	0	.
L * H	Sweden		30°C	0	.
F * P		high	control	-0.5464	1.6097
F * P		high	16 days	1.9350	1.6923
F * P		high	complete F0	0	.
F * P		low	control	0	.
F * P		low	16 days	0	.
F * P		low	complete F0	0	.
F * H		high	20°C	-1.1325	1.7449
F * H		high	30°C	0	.
F * H		low	20°C	0	.
F * H		low	30°C	0	.
P * H			control	3.4063	1.7169
P * H			control	0	.

P * H			16 days	20°C	3.1795	1.5874
P * H			16 days	30°C	0	.
P * H			complete F0	20°C	0	.
P * H			complete F0	30°C	0	.
L * F * P	France	high	control		0.3297	1.7939
L * F * P	France	high	16 days		-1.9363	1.8601
L * F * P	France	high	complete F0		0	.
L * F * P	France	low	control		0	.
L * F * P	France	low	16 days		0	.
L * F * P	France	low	complete F0		0	.
L * F * P	Sweden	high	control		0	.
L * F * P	Sweden	high	16 days		0	.
L * F * P	Sweden	high	complete F0		0	.
L * F * P	Sweden	low	control		0	.
L * F * P	Sweden	low	16 days		0	.
L * F * P	Sweden	low	complete F0		0	.
L * F * H	France	high		20°C	1.2245	2.0653
L * F * H	France	high		30°C	0	.
L * F * H	France	low		20°C	0	.
L * F * H	France	low		30°C	0	.
L * F * H	Sweden	high		20°C	0	.
L * F * H	Sweden	high		30°C	0	.
L * F * H	Sweden	low		20°C	0	.
L * F * H	Sweden	low		30°C	0	.
L * P * H	France		control	20°C	-3.6558	1.9738
L * P * H	France		control	30°C	0	.
L * P * H	France		16 days	20°C	-2.8637	1.8786
L * P * H	France		16 days	30°C	0	.
L * P * H	France		complete F0	20°C	0	.
L * P * H	France		complete F0	30°C	0	.

L * P * H	Sweden	control	20°C	0	.
L * P * H	Sweden	control	30°C	0	.
L * P * H	Sweden	16 days	20°C	0	.
L * P * H	Sweden	16 days	30°C	0	.
L * P * H	Sweden	complete F0	20°C	0	.
L * P * H	Sweden	complete F0	30°C	0	.
F * P * H		high	control	20°C	2.0441 2.3688
F * P * H		high	control	30°C	0 .
F * P * H		high	16 days	20°C	-0.9099 2.1739
F * P * H		high	16 days	30°C	0 .
F * P * H		high	complete F0	20°C	0 .
F * P * H		high	complete F0	30°C	0 .
F * P * H		low	control	20°C	0 .
F * P * H		low	control	30°C	0 .
F * P * H		low	16 days	20°C	0 .
F * P * H		low	16 days	30°C	0 .
F * P * H		low	complete F0	20°C	0 .
F * P * H		low	complete F0	30°C	0 .
L * F * P * H	France	high	control	20°C	-1.9994 2.7947
L * F * P * H	France	high	control	30°C	0 .
L * F * P * H	France	high	16 days	20°C	0.8865 2.6710
L * F * P * H	France	high	16 days	30°C	0 .
.L * F * P * H	France	high	complete F0	20°C	0 .
L * F * P * H	France	high	complete F0	30°C	0 .
L * F * P * H	France	low	control	20°C	0 .
L * F * P * H	France	low	control	30°C	0 .
L * F * P * H	France	low	16 days	20°C	0 .
L * F * P * H	France	low	16 days	30°C	0 .
L * F * P * H	France	low	complete F0	20°C	0 .
L * F * P * H	France	low	complete F0	30°C	0 .

	L * F * P * H	Sweden	high	control	20°C	0	.
	L * F * P * H	Sweden	high	control	30°C	0	.
	L * F * P * H	Sweden	high	16 days	20°C	0	.
	L * F * P * H	Sweden	high	16 days	30°C	0	.
	L * F * P * H	Sweden	high	complete F0	20°C	0	.
	L * F * P * H	Sweden	high	complete F0	30°C	0	.
	L * F * P * H	Sweden	low	control	20°C	0	.
	L * F * P * H	Sweden	low	control	30°C	0	.
	L * F * P * H	Sweden	low	16 days	20°C	0	.
	L * F * P * H	Sweden	low	16 days	30°C	0	.
	L * F * P * H	Sweden	low	complete F0	20°C	0	.
	L * F * P * H	Sweden	low	complete F0	30°C	0	.
Development time	Sex	female				0.7022	0.8827
	Sex	male				0	.
	Latitude (L)	France				-39.8475	2.2719
	Latitude (L)	Sweden				0	.
	Food (F)		high			4.2302	2.6304
	Food (F)		low			0	.
	Pesticide (P)			control		7.1435	2.5745
	Pesticide (P)			16 days		4.4214	2.6512
	Pesticide (P)			complete F0		0	.
	L * F	France	high			-7.8675	3.22865
	L * F	France	low			0	.
	L * F	Sweden	high			0	.
	L * F	Sweden	low			0	.
	L * P	France		control		-6.4352	3.1724
	L * P	France		16 days		-4.4343	3.2337
	L * P	France		complete F0		0	.
	L * P	Sweden		control		0	.

	L * P	Sweden	16 days	0	.	
	L * P	Sweden	complete F0	0	.	
	F * P	high	control	-14.1857	3.6077	
	F * P	high	16 days	-10.0882	3.6722	
	F * P	high	complete F0	0	.	
	F * P	low	control	0	.	
	F * P	low	16 days	0	.	
	F * P	low	complete F0	0	.	
	L * F * P	France	high	control	12.6929	4.4734
	L * F * P	France	high	16 days	9.5663	4.5321
	L * F * P	France	high	complete F0	0	.
	L * F * P	France	low	control	0	.
	L * F * P	France	low	16 days	0	.
	L * F * P	France	low	complete F0	0	.
	L * F * P	Sweden	high	control	0	.
	L * F * P	Sweden	high	16 days	0	.
	L * F * P	Sweden	high	complete F0	0	.
	L * F * P	Sweden	low	control	0	.
	L * F * P	Sweden	low	16 days	0	.
	L * F * P	Sweden	low	complete F0	0	.
Dry mass	Sex	female		0.1664	0.02774	
	Sex	male		0	.	
	Latitude (L)	France		-0.6463	0.1643	
	Latitude (L)	Sweden		0	.	
	Food (F)	high		0.2687	0.1019	
	Food (F)	low		0	.	
	Pesticide (P)		control	-0.01375	0.09780	
	Pesticide (P)		16 days	0.04344	0.1010	
	Pesticide (P)		complete F0	0	.	

Heat (H)				20°C	0.02070	0.1039
Heat (H)				30°C	0	.
L * F	France	high			0.1283	0.1346
L * F	France	low			0	.
L * F	Sweden	high			0	.
L * F	Sweden	low			0	.
L * P	France		control		0.03015	0.1309
L * P	France		16 days		0.01611	0.1332
L * P	France		complete F0		0	.
L * P	Sweden		control		0	.
L * P	Sweden		16 days		0	.
L * P	Sweden		complete F0		0	.
L * H	France			20°C	0.04574	0.1355
L * H	France			30°C	0	.
L * H	Sweden			20°C	0	.
L * H	Sweden			30°C	0	.
F * P		high	control		0.01134	0.1389
F * P		high	16 days		-0.2112	0.1451
F * P		high	complete F0		0	.
F * P		low	control		0	.
F * P		low	16 days		0	.
F * P		low	complete F0		0	.
F * H		high		20°C	-0.1215	0.1504
F * H		high		30°C	0	.
F * H		low		20°C	0	.
F * H		low		30°C	0	.
P * H			control	20°C	0.1058	0.1427
P * H			control	30°C	0	.
P * H			16 days	20°C	0.02412	0.1457
P * H			16 days	30°C	0	.

P * H			complete F0	20°C	0	.
P * H			complete F0	30°C	0	.
L * F * P	France	high	control		0.1767	0.1870
L * F * P	France	high	16 days		0.1828	0.1916
L * F * P	France	high	complete F0		0	.
L * F * P	France	low	control		0	.
L * F * P	France	low	16 days		0	.
L * F * P	France	low	complete F0		0	.
L * F * P	Sweden	high	control		0	.
L * F * P	Sweden	high	16 days		0	.
L * F * P	Sweden	high	complete F0		0	.
L * F * P	Sweden	low	control		0	.
L * F * P	Sweden	low	16 days		0	.
L * F * P	Sweden	low	complete F0		0	.
L * F * H	France	high		20°C	0.1629	0.1951
L * F * H	France	high		30°C	0	.
L * F * H	France	low		20°C	0	.
L * F * H	France	low		30°C	0	.
L * F * H	Sweden	high		20°C	0	.
L * F * H	Sweden	high		30°C	0	.
L * F * H	Sweden	low		20°C	0	.
L * F * H	Sweden	low		30°C	0	.
L * P * H	France		control	20°C	0.01017	0.1881
L * P * H	France		control	30°C	0	.
L * P * H	France		16 days	20°C	-0.04670	0.1911
L * P * H	France		16 days	30°C	0	.
L * P * H	France		complete F0	20°C	0	.
L * P * H	France		complete F0	30°C	0	.
L * P * H	Sweden		control	20°C	0	.
L * P * H	Sweden		control	30°C	0	.

L * P * H	Sweden		16 days	20°C	0	.
L * P * H	Sweden		16 days	30°C	0	.
L * P * H	Sweden		complete F0	20°C	0	.
L * P * H	Sweden		complete F0	30°C	0	.
F * P * H		high	control	20°C	-0.00037	0.2031
F * P * H		high	control	30°C	0	.
F * P * H		high	16 days	20°C	0.2163	0.2077
F * P * H		high	16 days	30°C	0	.
F * P * H		high	complete F0	20°C	0	.
F * P * H		high	complete F0	30°C	0	.
F * P * H		low	control	20°C	0	.
F * P * H		low	control	30°C	0	.
F * P * H		low	16 days	20°C	0	.
F * P * H		low	16 days	30°C	0	.
F * P * H		low	complete F0	20°C	0	.
F * P * H		low	complete F0	30°C	0	.
L * F * P * H	France	high	control	20°C	-0.3000	0.2681
L * F * P * H	France	high	control	30°C	0	.
L * F * P * H	France	high	16 days	20°C	-0.2729	0.2725
L * F * P * H	France	high	16 days	30°C	0	.
L * F * P * H	France	high	complete F0	20°C	0	.
L * F * P * H	France	high	complete F0	30°C	0	.
L * F * P * H	France	low	control	20°C	0	.
L * F * P * H	France	low	control	30°C	0	.
L * F * P * H	France	low	16 days	20°C	0	.
L * F * P * H	France	low	16 days	30°C	0	.
L * F * P * H	France	low	complete F0	20°C	0	.
L * F * P * H	France	low	complete F0	30°C	0	.
L * F * P * H	Sweden	high	control	20°C	0	.
L * F * P * H	Sweden	high	control	30°C	0	.

	L * F * P * H	Sweden	high	16 days	20°C	0	.
	L * F * P * H	Sweden	high	16 days	30°C	0	.
	L * F * P * H	Sweden	high	complete F0	20°C	0	.
	L * F * P * H	Sweden	high	complete F0	30°C	0	.
	L * F * P * H	Sweden	low	control	20°C	0	.
	L * F * P * H	Sweden	low	control	30°C	0	.
	L * F * P * H	Sweden	low	16 days	20°C	0	.
	L * F * P * H	Sweden	low	16 days	30°C	0	.
	L * F * P * H	Sweden	low	complete F0	20°C	0	.
	L * F * P * H	Sweden	low	complete F0	30°C	0	.
Water content	Sex	female				0.1763	0.06624
	Sex	male				0	.
	Latitude (L)	France				0.7253	0.2508
	Latitude (L)	Sweden				0	.
	Food (F)		high			0.1725	0.2425
	Food (F)		low			0	.
	Pesticide (P)			control		-0.1158	0.2332
	Pesticide (P)			16 days		-0.6174	0.2408
	Pesticide (P)			complete F0		0	.
	Heat (H)				20°C	0.3922	0.2474
	Heat (H)				30°C	0	.
	L * F	France	high			-0.5717	0.3207
	L * F	France	low			0	.
	L * F	Sweden	high			0	.
	L * F	Sweden	low			0	.
	L * P	France		control		0.04289	0.3123
	L * P	France		16 days		0.5954	0.3178
	L * P	France		complete F0		0	.
	L * P	Sweden		control		0	.

L * P	Sweden		16 days		0	.
L * P	Sweden		complete F0		0	.
L * H	France			20°C	-0.05808	0.3228
L * H	France			30°C	0	.
L * H	Sweden			20°C	0	.
L * H	Sweden			30°C	0	.
F * P		high	control		-0.2526	0.3311
F * P		high	16 days		0.3095	0.3453
F * P		high	complete F0		0	.
F * P		low	control		0	.
F * P		low	16 days		0	.
F * P		low	complete F0		0	.
F * H		high		20°C	-0.2641	0.3582
F * H		high		30°C	0	.
F * H		low		20°C	0	.
F * H		low		30°C	0	.
P * H			control	20°C	0.03193	0.3422
P * H			control	30°C	0	.
P * H			16 days	20°C	0.7001	0.3503
P * H			16 days	30°C	0	.
P * H			complete F0	20°C	0	.
P * H			complete F0	30°C	0	.
L * F * P	France	high	control		-0.00554	0.4460
L * F * P	France	high	16 days		0	.
L * F * P	France	high	complete F0		-0.4651	0.4566
L * F * P	France	low	control		0	.
L * F * P	France	low	16 days		0	.
L * F * P	France	low	complete F0		0	.
L * F * P	Sweden	high	control		0	.
L * F * P	Sweden	high	16 days		0	.

L * F * P	Sweden	high	complete F0		0	.
L * F * P	Sweden	low	control		0	.
L * F * P	Sweden	low	16 days		0	.
L * F * P	Sweden	low	complete F0		0	.
L * F * H	France	high		20°C	0.2544	0.4649
L * F * H	France	high		30°C	0	.
L * F * H	France	low		20°C	0	.
L * F * H	France	low		30°C	0	.
L * F * H	Sweden	high		20°C	0	.
L * F * H	Sweden	high		30°C	0	.
L * F * H	Sweden	low		20°C	0	.
L * F * H	Sweden	low		30°C	0	.
L * P * H	France		control	20°C	-0.1374	0.4502
L * P * H	France		control	30°C	0	.
L * P * H	France		16 days	20°C	-0.8883	0.4579
L * P * H	France		16 days	30°C	0	.
L * P * H	France		complete F0	20°C	0	.
L * P * H	France		complete F0	30°C	0	.
L * P * H	Sweden		control	20°C	0	.
L * P * H	Sweden		control	30°C	0	.
L * P * H	Sweden		16 days	20°C	0	.
L * P * H	Sweden		16 days	30°C	0	.
L * P * H	Sweden		complete F0	20°C	0	.
L * P * H	Sweden		complete F0	30°C	0	.
F * P * H		high	control	20°C	0.09846	0.4859
F * P * H		high	control	30°C	0	.
F * P * H		high	16 days	20°C	-0.4577	0.4987
F * P * H		high	16 days	30°C	0	.
F * P * H		high	complete F0	20°C	0	.
F * P * H		high	complete F0	30°C	0	.

F * P * H		low	control	20°C	0	.
F * P * H		low	control	30°C	0	.
F * P * H		low	16 days	20°C	0	.
F * P * H		low	16 days	30°C	0	.
F * P * H		low	complete F0	20°C	0	.
F * P * H		low	complete F0	30°C	0	.
L * F * P * H	France	high	control	20°C	0.3139	0.6407
L * F * P * H	France	high	control	30°C	0	.
L * F * P * H	France	high	16 days	20°C	0.7531	0.6524
L * F * P * H	France	high	16 days	30°C	0	.
L * F * P * H	France	high	complete F0	20°C	0	.
L * F * P * H	France	high	complete F0	30°C	0	.
L * F * P * H	France	low	control	20°C	0	.
L * F * P * H	France	low	control	30°C	0	.
L * F * P * H	France	low	16 days	20°C	0	.
L * F * P * H	France	low	16 days	30°C	0	.
L * F * P * H	France	low	complete F0	20°C	0	.
L * F * P * H	France	low	complete F0	30°C	0	.
L * F * P * H	Sweden	high	control	20°C	0	.
L * F * P * H	Sweden	high	control	30°C	0	.
L * F * P * H	Sweden	high	16 days	20°C	0	.
L * F * P * H	Sweden	high	16 days	30°C	0	.
L * F * P * H	Sweden	high	complete F0	20°C	0	.
L * F * P * H	Sweden	high	complete F0	30°C	0	.
L * F * P * H	Sweden	low	control	20°C	0	.
L * F * P * H	Sweden	low	control	30°C	0	.
L * F * P * H	Sweden	low	16 days	20°C	0	.
L * F * P * H	Sweden	low	16 days	30°C	0	.
L * F * P * H	Sweden	low	complete F0	20°C	0	.
L * F * P * H	Sweden	low	complete F0	30°C	0	.

Fat content	Sex	female				-0.1374	0.1118
	Sex	male				0	.
	Latitude (L)		France			-1.8754	0.3939
	Latitude (L)		Sweden			0	.
	Food (F)			high		1.0003	0.4471
	Food (F)			low		0	.
	Pesticide (P)				control	0.1377	0.4257
	Pesticide (P)				16 days	-0.7559	0.4180
	Pesticide (P)				complete F0	0	.
	Heat (H)					20°C	0.09057
	Heat (H)					30°C	0
	L * F		France	high			0.5809
	L * F		France	low			0
	L * F		Sweden	high			0
	L * F		Sweden	low			0
	L * P		France		control		0.1539
	L * P		France		16 days		0.8839
	L * P		France		complete F0		0
	L * P		Sweden		control		0
	L * P		Sweden		16 days		0
	L * P		Sweden		complete F0		0
	L * H		France			20°C	1.0166
	L * H		France			30°C	0
	L * H		Sweden			20°C	0
	L * H		Sweden			30°C	0
	F * P			high	control		-0.17194
	F * P			high	16 days		0.5620
	F * P			high	complete F0		0
	F * P			low	control		0

F * P		low	16 days		0	.
F * P		low	complete F0		0	.
F * H		high		20°C	-0.2412	0.6475
F * H		high		30°C	0	.
F * H		low		20°C	0	.
F * H		low		30°C	0	.
P * H			control	20°C	0.2031	0.4935
P * H			control	30°C	0	.
P * H			16 days	20°C	0.9843	0.4935
P * H			16 days	30°C	0	.
P * H			complete F0	20°C	0	.
P * H			complete F0	30°C	0	.
L * F * P	France	high	control		0.2943	0.7831
L * F * P	France	high	16 days		-0.6110	0.7874
L * F * P	France	high	complete F0		0	.
L * F * P	France	low	control		0	.
L * F * P	France	low	16 days		0	.
L * F * P	France	low	complete F0		0	.
L * F * P	Sweden	high	control		0	.
L * F * P	Sweden	high	16 days		0	.
L * F * P	Sweden	high	complete F0		0	.
L * F * P	Sweden	low	control		0	.
L * F * P	Sweden	low	16 days		0	.
L * F * P	Sweden	low	complete F0		0	.
L * F * H	France	high		20°C	-0.1882	0.8114
L * F * H	France	high		30°C	0	.
L * F * H	France	low		20°C	0	.
L * F * H	France	low		30°C	0	.
L * F * H	Sweden	high		20°C	0	.
L * F * H	Sweden	high		30°C	0	.

L * F * H	Sweden	low		20°C	0	.
L * F * H	Sweden	low		30°C	0	.
L * P * H	France		control	20°C	-0.5483	0.7656
L * P * H	France		control	30°C	0	.
L * P * H	France		16 days	20°C	-1.1392	0.7670
L * P * H	France		16 days	30°C	0	.
L * P * H	France		complete F0	20°C	0	.
L * P * H	France		complete F0	30°C	0	.
L * P * H	Sweden		control	20°C	0	.
L * P * H	Sweden		control	30°C	0	.
L * P * H	Sweden		16 days	20°C	0	.
L * P * H	Sweden		16 days	30°C	0	.
L * P * H	Sweden		complete F0	20°C	0	.
L * P * H	Sweden		complete F0	30°C	0	.
F * P * H		high	control	20°C	0.2630	0.8625
F * P * H		high	control	30°C	0	.
F * P * H		high	16 days	20°C	0.08102	0.8686
F * P * H		high	16 days	30°C	0	.
F * P * H		high	complete F0	20°C	0	.
F * P * H		high	complete F0	30°C	0	.
F * P * H		low	control	20°C	0	.
F * P * H		low	control	30°C	0	.
F * P * H		low	16 days	20°C	0	.
F * P * H		low	16 days	30°C	0	.
F * P * H		low	complete F0	20°C	0	.
F * P * H		low	complete F0	30°C	0	.
L * F * P * H	France	high	control	20°C	-0.08763	1.1049
L * F * P * H	France	high	control	30°C	0	.
L * F * P * H	France	high	16 days	20°C	-0.1268	1.1122
L * F * P * H	France	high	16 days	30°C	0	.

L * F * P * H	France	high	complete F0	20°C	0	.
L * F * P * H	France	high	complete F0	30°C	0	.
L * F * P * H	France	low	control	20°C	0	.
L * F * P * H	France	low	control	30°C	0	.
L * F * P * H	France	low	16 days	20°C	0	.
L * F * P * H	France	low	16 days	30°C	0	.
L * F * P * H	France	low	complete F0	20°C	0	.
L * F * P * H	France	low	complete F0	30°C	0	.
L * F * P * H	Sweden	high	control	20°C	0	.
L * F * P * H	Sweden	high	control	30°C	0	.
L * F * P * H	Sweden	high	16 days	20°C	0	.
L * F * P * H	Sweden	high	16 days	30°C	0	.
L * F * P * H	Sweden	high	complete F0	20°C	0	.
L * F * P * H	Sweden	high	complete F0	30°C	0	.
L * F * P * H	Sweden	low	control	20°C	0	.
L * F * P * H	Sweden	low	control	30°C	0	.
L * F * P * H	Sweden	low	16 days	20°C	0	.
L * F * P * H	Sweden	low	16 days	30°C	0	.
L * F * P * H	Sweden	low	complete F0	20°C	0	.
L * F * P * H	Sweden	low	complete F0	30°C	0	.

Hsp70 levels	Sex	female			-0.1563	0.0963
	Sex	male			0	.
	Latitude (L)		France		-1.2342	0.3346
	Latitude (L)		Sweden		0	.
	Food (F)			high	1.7707	0.3286
	Food (F)			low	0	.
	Pesticide (P)			control	0.1167	0.3346
	Pesticide (P)			16 days	0.1298	0.3344
	Pesticide (P)			complete F0	0	.

Heat (H)				20°C	-0.03578	0.3283
Heat (H)				30°C	0	.
L * F	France	high			-0.6891	0.4590
L * F	France	low			0	.
L * F	Sweden	high			0	.
L * F	Sweden	low			0	.
L * P	France		control		-0.5536	0.4679
L * P	France		16 days		-0.2715	0.4677
L * P	France		complete F0		0	.
L * P	Sweden		control		0	.
L * P	Sweden		16 days		0	.
L * P	Sweden		complete F0		0	.
L * H	France			20°C	-0.1543	0.4634
L * H	France			30°C	0	.
L * H	Sweden			20°C	0	.
L * H	Sweden			30°C	0	.
F * P		high	control		-0.7994	0.4589
F * P		high	16 days		0.1423	0.4687
F * P		high	complete F0		0	.
F * P		low	control		0	.
F * P		low	16 days		0	.
F * P		low	complete F0		0	.
F * H		high		20°C	-0.9188	0.4649
F * H		high		30°C	0	.
F * H		low		20°C	0	.
F * H		low		30°C	0	.
P * H			control	20°C	-1.0935	0.4687
P * H			control	30°C	0	.
P * H			16 days	20°C	-0.7499	0.4636
P * H			16 days	30°C	0	.

P * H			complete F0	20°C	0	.
P * H			complete F0	30°C	0	.
L * F * P	France	high	control		0.6761	0.6521
L * F * P	France	high	16 days		-0.1333	0.6589
L * F * P	France	high	complete F0		0	.
L * F * P	France	low	control		0	.
L * F * P	France	low	16 days		0	.
L * F * P	France	low	complete F0		0	.
L * F * P	Sweden	high	control		0	.
L * F * P	Sweden	high	16 days		0	.
L * F * P	Sweden	high	complete F0		0	.
L * F * P	Sweden	low	control		0	.
L * F * P	Sweden	low	16 days		0	.
L * F * P	Sweden	low	complete F0		0	.
L * F * H	France	high		20°C	0.4769	0.6521
L * F * H	France	high		30°C	0	.
L * F * H	France	low		20°C	0	.
L * F * H	France	low		30°C	0	.
L * F * H	Sweden	high		20°C	0	.
L * F * H	Sweden	high		30°C	0	.
L * F * H	Sweden	low		20°C	0	.
L * F * H	Sweden	low		30°C	0	.
L * P * H	France		control	20°C	1.0826	0.6621
L * P * H	France		control	30°C	0	.
L * P * H	France		16 days	20°C	0.9435	0.6516
L * P * H	France		16 days	30°C	0	.
L * P * H	France		complete F0	20°C	0	.
L * P * H	France		complete F0	30°C	0	.
L * P * H	Sweden		control	20°C	0	.
L * P * H	Sweden		control	30°C	0	.

L * P * H	Sweden		16 days	20°C	0	.
L * P * H	Sweden		16 days	30°C	0	.
L * P * H	Sweden		complete F0	20°C	0	.
L * P * H	Sweden		complete F0	30°C	0	.
F * P * H		high	control	20°C	0.006498	0.6559
F * P * H		high	control	30°C	0	.
F * P * H		high	16 days	20°C	0.6848	0.6567
F * P * H		high	16 days	30°C	0	.
F * P * H		high	complete F0	20°C	0	.
F * P * H		high	complete F0	30°C	0	.
F * P * H		low	control	20°C	0	.
F * P * H		low	control	30°C	0	.
F * P * H		low	16 days	20°C	0	.
F * P * H		low	16 days	30°C	0	.
F * P * H		low	complete F0	20°C	0	.
F * P * H		low	complete F0	30°C	0	.
L * F * P * H	France	high	control	20°C	-0.02101	0.9245
L * F * P * H	France	high	control	30°C	0	.
L * F * P * H	France	high	16 days	20°C	-0.6629	0.9245
L * F * P * H	France	high	16 days	30°C	0	.
L * F * P * H	France	high	complete F0	20°C	0	.
L * F * P * H	France	high	complete F0	30°C	0	.
L * F * P * H	France	low	control	20°C	0	.
L * F * P * H	France	low	control	30°C	0	.
L * F * P * H	France	low	16 days	20°C	0	.
L * F * P * H	France	low	16 days	30°C	0	.
L * F * P * H	France	low	complete F0	20°C	0	.
L * F * P * H	France	low	complete F0	30°C	0	.
L * F * P * H	Sweden	high	control	20°C	0	.
L * F * P * H	Sweden	high	control	30°C	0	.

			L * F * P * H	Sweden	high	16 days	20°C	0	.
			L * F * P * H	Sweden	high	16 days	30°C	0	.
			L * F * P * H	Sweden	high	complete F0	20°C	0	.
			L * F * P * H	Sweden	high	complete F0	30°C	0	.
			L * F * P * H	Sweden	low	control	20°C	0	.
			L * F * P * H	Sweden	low	control	30°C	0	.
			L * F * P * H	Sweden	low	16 days	20°C	0	.
			L * F * P * H	Sweden	low	16 days	30°C	0	.
			L * F * P * H	Sweden	low	complete F0	20°C	0	.
			L * F * P * H	Sweden	low	complete F0	30°C	0	.
PO activity	protein							9.7817	5.0519
	mass							0.04709	0.07243
	Sex	female						-0.09805	0.3439
	Sex	male						0	.
	Latitude (L)			France				-1.5662	1.1323
	Latitude (L)			Sweden				0	.
	Food (F)				high			5.6592	1.1971
	Food (F)				low			0	.
	Pesticide (P)					control		6.1311	1.1525
	Pesticide (P)					16 days		2.4749	1.1937
	Pesticide (P)					complete F0		0	.
	Heat (H)						20°C	1.2363	1.2215
	Heat (H)						30°C	0	.
	L * F			France	high			-4.5605	1.5807
	L * F			France	low			0	.
	L * F			Sweden	high			0	.
	L * F			Sweden	low			0	.
	L * P			France		control		-2.8720	1.5438
	L * P			France		16 days		0.09912	1.5739

L * P	France		complete F0	0	.
L * P	Sweden		control	0	.
L * P	Sweden		16 days	0	.
L * P	Sweden		complete F0	0	.
L * H	France		20°C	1.5497	1.0996
L * H	France		30°C	0	.
L * H	Sweden		20°C	0	.
L * H	Sweden		30°C	0	.
F * P		high	control	-3.2476	1.6356
F * P		high	16 days	-1.0470	1.6962
F * P		high	complete F0	0	.
F * P		low	control	0	.
F * P		low	16 days	0	.
F * P		low	complete F0	0	.
F * H		high	20°C	-1.1940	1.2679
F * H		high	30°C	0	.
F * H		low	20°C	0	.
F * H		low	30°C	0	.
P * H			control 20°C	1.0930	1.6860
P * H			control 30°C	0	.
P * H			16 days 20°C	2.6207	1.7311
P * H			16 days 30°C	0	.
P * H			complete F0 20°C	0	.
P * H			complete F0 30°C	0	.
L * F * P	France	high	control	1.8043	2.2065
L * F * P	France	high	16 days	-0.4585	2.2499
L * F * P	France	high	complete F0	0	.
L * F * P	France	low	control	0	.
L * F * P	France	low	16 days	0	.
L * F * P	France	low	complete F0	0	.

L * F * P	Sweden	high	control		0	.
L * F * P	Sweden	high	16 days		0	.
L * F * P	Sweden	high	complete F0		0	.
L * F * P	Sweden	low	control		0	.
L * F * P	Sweden	low	16 days		0	.
L * F * P	Sweden	low	complete F0		0	.
L * F * H	France	high		20°C	0.1803	2.2974
L * F * H	France	high		30°C	0	.
L * F * H	France	low		20°C	0	.
L * F * H	France	low		30°C	0	.
L * F * H	Sweden	high		20°C	0	.
L * F * H	Sweden	high		30°C	0	.
L * F * H	Sweden	low		20°C	0	.
L * F * H	Sweden	low		30°C	0	.
L * P * H	France		control	20°C	-4.0184	2.2220
L * P * H	France		control	30°C	0	.
L * P * H	France		16 days	20°C	0.6042	2.2646
L * P * H	France		16 days	30°C	0	.
L * P * H	France		complete F0	20°C	0	.
L * P * H	France		complete F0	30°C	0	.
L * P * H	Sweden		control	20°C	0	.
L * P * H	Sweden		control	30°C	0	.
L * P * H	Sweden		16 days	20°C	0	.
L * P * H	Sweden		16 days	30°C	0	.
L * P * H	Sweden		complete F0	20°C	0	.
L * P * H	Sweden		complete F0	30°C	0	.
F * P * H		high	control	20°C	1.779	2.4002
F * P * H		high	control	30°C	0	.
F * P * H		high	16 days	20°C	4.7315	2.4501
F * P * H		high	16 days	30°C	0	.

F * P * H		high	complete F0	20°C	0	.
F * P * H		high	complete F0	30°C	0	.
F * P * H		low	control	20°C	0	.
F * P * H		low	control	30°C	0	.
F * P * H		low	16 days	20°C	0	.
F * P * H		low	16 days	30°C	0	.
F * P * H		low	complete F0	20°C	0	.
F * P * H		low	complete F0	30°C	0	.
L * F * P * H	France	high	control	20°C	3.6195	3.1668
L * F * P * H	France	high	control	30°C	0	.
L * F * P * H	France	high	16 days	20°C	-2.6111	3.2178
L * F * P * H	France	high	16 days	30°C	0	.
L * F * P * H	France	high	complete F0	20°C	0	.
L * F * P * H	France	high	complete F0	30°C	0	.
L * F * P * H	France	low	control	20°C	0	.
L * F * P * H	France	low	control	30°C	0	.
L * F * P * H	France	low	16 days	20°C	0	.
L * F * P * H	France	low	16 days	30°C	0	.
L * F * P * H	France	low	complete F0	20°C	0	.
L * F * P * H	France	low	complete F0	30°C	0	.
L * F * P * H	Sweden	high	control	20°C	0	.
L * F * P * H	Sweden	high	control	30°C	0	.
L * F * P * H	Sweden	high	16 days	20°C	0	.
L * F * P * H	Sweden	high	16 days	30°C	0	.
L * F * P * H	Sweden	high	complete F0	20°C	0	.
L * F * P * H	Sweden	high	complete F0	30°C	0	.
L * F * P * H	Sweden	low	control	20°C	0	.
L * F * P * H	Sweden	low	control	30°C	0	.
L * F * P * H	Sweden	low	16 days	20°C	0	.
L * F * P * H	Sweden	low	16 days	30°C	0	.

712	L * F * P * H	Sweden	low	complete F0	20°C	0	.
	L * F * P * H	Sweden	low	complete F0	30°C	0	.
713							

714 Appendix 2. Full model results of the ANOVAs testing for effects of latitude, food level,
715 pesticide concentration and adult heat condition on the response variables under study.

Response variable	Effect	df 1, df 2	<i>F</i>	<i>P</i>
Survival	Latitude (L)	1, 4	76.72	< 0.001
	Food (F)	1, 792	0.41	0.52
	Pesticide (P)	2, 792	0.37	0.69
	Heat (H)	1, 792	0.24	0.62
	L x F	1, 792	0.15	0.70
	L x P	2, 792	0.32	0.72
	L x H	1, 792	0.060	0.80
	F x P	2, 792	1.69	0.19
	F x H	1, 792	0.29	0.59
	P x H	2, 792	0.93	0.39
	L x F x P	2, 792	1.45	0.24
	L x F x H	1, 792	2.20	0.14
	L x P x H	2, 792	1.06	0.35
	F x P x H	2, 792	0.70	0.50
	L x F x P x H	2, 792	0.14	0.87
Flying ability	Sex	1, 554	4.34	0.038
	Latitude (L)	1, 4	0.050	0.83
	Food (F)	1, 554	0.090	0.76
	Pesticide (P)	2, 554	0.33	0.72
	Heat (H)	1, 554	28.31	< 0.001
	L x F	1, 554	0.34	0.56
	L x P	2, 554	0.070	0.94
	L x H	1, 554	0.35	0.56
	F x P	2, 554	0.78	0.46
	F x H	1, 554	0.90	0.34
	P x H	2, 554	1.19	0.25
	L x F x P	2, 554	0.80	0.45
	L x F x H	1, 554	0.32	0.57
	L x P x H	2, 554	1.19	0.25
	F x P x H	2, 554	1.00	0.37
	L x F x P x H	2, 554	0.44	0.64
Development time	Sex	1, 605	0.63	0.43
	Latitude (L)	1, 605	2288.39	< 0.001
	Food (F)	1, 605	20.01	< 0.001
	Pesticide (P)	2, 605	0.11	0.90
	L x F	1, 605	0.060	0.81
	L x P	2, 605	0.020	0.98
	F x P	2, 605	6.35	0.0019
	L x F x P	2, 605	4.32	0.014
Dry mass	Sex	1, 441	36.00	< 0.001
	Latitude (L)	1, 5.85	12.82	0.012
	Food (F)	1, 441	115.87	< 0.001
	Pesticide (P)	2, 441	2.47	0.086

	Heat (H)	1, 442	3.17	0.076
	L x F	1, 441	18.32	< 0.001
	L x P	2, 441	0.28	0.76
	L x H	1, 442	0.13	0.72
	F x P	2, 441	1.35	0.26
	F x H	1, 441	1.35	0.25
	P x H	2, 441	0.21	0.81
	L x F x P	2, 441	0.060	0.94
	L x F x H	1, 441	0.070	0.80
	L x P x H	2, 441	0.99	0.37
	F x P x H	2, 441	1.57	0.21
	L x F x P x H	2, 441	0.75	0.47
Water content	Sex	1, 441	7.08	0.0081
	Latitude (L)	1, 6.27	14.60	0.0080
	Food (F)	1, 439	10.51	0.0013
	Pesticide (P)	2, 439	3.11	0.046
	Heat (H)	1, 441	36.43	< 0.001
	L x F	1, 439	10.47	0.0013
	L x P	2, 439	0.22	0.81
	L x H	1, 441	0.52	0.47
	F x P	2, 439	0.59	0.55
	F x H	1, 439	0.36	0.55
	P x H	2, 439	0.89	0.41
	L x F x P	2, 439	0.29	0.75
	L x F x H	1, 439	1.43	0.20
	L x P x H	2, 439	1.76	0.17
	F x P x H	2, 440	0.62	0.54
	L x F x P x H	2, 440	0.67	0.51
Fat content	Sex	1, 423	1.51	0.22
	Latitude (L)	1, 4.2	49.30	0.0018
	Food (F)	1, 419	130.37	< 0.001
	Pesticide (P)	2, 421	2.75	0.065
	Heat (H)	1, 423	28.38	< 0.001
	L x F	1, 419	2.40	0.12
	L x P	2, 421	0.010	0.99
	L x H	1, 423	2.11	0.15
	F x P	2, 420	3.48	0.062
	F x H	1, 420	1.32	0.25
	P x H	2, 422	2.36	0.076
	L x F x P	2, 420	1.60	0.20
	L x F x H	1, 420	0.34	0.56
	L x P x H	2, 421	1.40	0.25
	F x P x H	2, 424	0.10	0.90
	L x F x P x H	2; 424	0.010	0.99
Hsp70 levels	Sex	1, 287	2.63	0.11
	Latitude (L)	1, 287	231.15	< 0.001
	Food (F)	1, 287	114.86	< 0.001

	Pesticide (P)	2, 287	21.29	< 0.001
	Heat (H)	1, 287	50.07	< 0.001
	L x F	1, 287	4.09	0.44
	L x P	2, 287	1.39	0.25
	L x H	1, 287	11.59	< 0.001
	F x P	2, 287	4.85	0.0085
	F x H	1, 287	8.84	0.0032
	P x H	2, 287	3.20	0.042
	L x F x P	2, 287	2.47	0.083
	L x F x H	1, 287	0.43	0.51
	L x P x H	2, 287	1.68	0.21
	F x P x H	2, 287	0.39	0.69
	L x F x P x H	2, 287	0.33	0.72
PO activity	Protein	1, 444	3.75	0.054
	Mass	1, 444	0.42	0.52
	Sex	1, 444	0.080	0.78
	Latitude (L)	1, 444	107.74	< 0.001
	Food (F)	1, 444	52.48	< 0.001
	Pesticide (P)	2, 444	57.99	< 0.001
	Heat (H)	1, 444	1.03	0.31
	L x F	1, 444	1.30	0.26
	L x P	2, 444	1.10	0.29
	L x H	1, 444	34.63	< 0.001
	F x P	2, 444	1.40	0.25
	F x H	1, 444	20.53	< 0.001
	P x H	2, 444	1.84	0.17
	L x F x P	2, 444	6.14	0.0023
	L x F x H	1, 444	0.16	0.69
	L x P x H	2, 444	1.03	0.36
	F x P x H	2, 444	3.37	0.035
	L x F x P x H	2, 444	1.98	0.14

716

717

718 Appendix 3. Means and standard error for each of the treatment combinations per response
719 variable.

Response variable	Latitude	Food	Pesticide	Heat	Mean	Standard error	N
Development time	France	high	control	20°C	21.46	1.33	69
	France	high	16 days	20°C	21.71	1.32	68
	France	high	complete F0	20°C	22.24	1.34	67
	France	low	control	20°C	26.59	1.32	67
	France	low	16 days	20°C	25.87	1.32	69
	France	low	complete F0	20°C	25.88	1.31	70
	Sweden	high	control	20°C	62.91	1.71	63
	Sweden	high	16 days	20°C	64.29	1.73	60
	Sweden	high	complete F0	20°C	69.96	1.86	60
	Sweden	low	control	20°C	72.87	1.78	57
	Sweden	low	16 days	20°C	70.15	1.89	51
	Sweden	low	complete F0	20°C	65.73	1.86	58
Dry mass	France	high	control	20°C	2.76	0.11	23
	France	high	control	30°C	2.84	0.12	21
	France	high	16 days	20°C	2.70	0.11	21
	France	high	16 days	30°C	2.67	0.12	20
	France	high	complete F0	20°C	2.74	0.11	21
	France	high	complete F0	30°C	2.64	0.12	21
	France	low	control	20°C	2.44	0.11	23
	France	low	control	30°C	2.26	0.11	21
	France	low	16 days	20°C	2.34	0.11	21
	France	low	16 days	30°C	2.30	0.11	21
	France	low	complete F0	20°C	2.31	0.11	21
	France	low	complete F0	30°C	2.24	0.11	22
	Sweden	high	control	20°C	3.16	0.12	16
	Sweden	high	control	30°C	3.15	0.12	19
	Sweden	high	16 days	20°C	3.13	0.12	18
	Sweden	high	16 days	30°C	2.99	0.12	16
	Sweden	high	complete F0	20°C	3.15	0.12	13
	Sweden	high	complete F0	30°C	3.05	0.12	15
	Sweden	low	control	20°C	3.00	0.12	17
	Sweden	low	control	30°C	2.87	0.12	17
	Sweden	low	16 days	20°C	2.97	0.12	16
	Sweden	low	16 days	30°C	2.92	0.12	15
	Sweden	low	complete F0	20°C	2.91	0.12	14
	Sweden	low	complete F0	30°C	2.89	0.12	17
Water content	France	high	control	20°C	89.04	0.17	23
	France	high	control	30°C	88.42	0.17	20
	France	high	16 days	20°C	89.00	0.17	21
	France	high	16 days	30°C	88.57	0.17	20
	France	high	complete F0	20°C	89.07	0.17	20
	France	high	complete F0	30°C	88.75	0.17	21
	France	low	control	20°C	89.30	0.17	23
	France	low	control	30°C	89.07	0.17	21

	France	low	16 days	20°C	89.27	0.17	21
	France	low	16 days	30°C	89.12	0.17	21
	France	low	complete F0	20°C	89.48	0.17	21
	France	low	complete F0	30°C	89.15	0.17	22
	Sweden	high	control	20°C	88.48	0.19	16
	Sweden	high	control	30°C	88.22	0.18	19
	Sweden	high	16 days	20°C	88.66	0.19	17
	Sweden	high	16 days	30°C	88.28	0.19	16
	Sweden	high	complete F0	20°C	88.72	0.21	13
	Sweden	high	complete F0	30°C	88.59	0.20	15
	Sweden	low	control	20°C	88.73	0.20	16
	Sweden	low	control	30°C	88.30	0.19	17
	Sweden	low	16 days	20°C	88.90	0.20	15
	Sweden	low	16 days	30°C	87.80	0.20	15
	Sweden	low	complete F0	20°C	88.81	0.20	14
	Sweden	low	complete F0	30°C	88.42	0.19	17
Fat content	France	high	control	20°C	9.89	0.25	22
	France	high	control	30°C	8.96	0.26	21
	France	high	16 days	20°C	9.00	0.26	21
	France	high	16 days	30°C	8.52	0.26	20
	France	high	complete F0	20°C	9.12	0.26	21
	France	high	complete F0	30°C	8.44	0.26	21
	France	low	control	20°C	8.15	0.25	23
	France	low	control	30°C	7.16	0.26	21
	France	low	16 days	20°C	7.94	0.26	21
	France	low	16 days	30°C	6.99	0.26	21
	France	low	complete F0	20°C	7.87	0.25	21
	France	low	complete F0	30°C	6.86	0.25	22
	Sweden	high	control	20°C	10.51	0.29	16
	Sweden	high	control	30°C	9.80	0.28	17
	Sweden	high	16 days	20°C	10.46	0.28	18
	Sweden	high	16 days	30°C	9.55	0.30	15
	Sweden	high	complete F0	20°C	9.74	0.35	10
	Sweden	high	complete F0	30°C	9.59	0.37	11
	Sweden	low	control	20°C	9.60	0.29	16
	Sweden	low	control	30°C	8.88	0.32	13
	Sweden	low	16 days	20°C	9.06	0.30	15
	Sweden	low	16 days	30°C	7.98	0.31	14
	Sweden	low	complete F0	20°C	8.83	0.31	14
	Sweden	low	complete F0	30°C	8.73	0.30	15
Hsp70 levels	France	high	control	20°C	7.99	0.23	12
	France	high	control	30°C	8.65	0.24	11
	France	high	16 days	20°C	8.66	0.23	12
	France	high	16 days	30°C	9.07	0.24	12
	France	high	complete F0	20°C	8.57	0.23	12
	France	high	complete F0	30°C	9.21	0.22	13
	France	low	control	20°C	7.49	0.24	11
	France	low	control	30°C	7.69	0.23	12

France	low	16 days	20°C	7.99	0.22	13
France	low	16 days	30°C	7.98	0.23	12
France	low	complete F0	20°C	7.93	0.23	12
France	low	complete F0	30°C	8.13	0.23	12
Sweden	high	control	20°C	8.40	0.23	12
Sweden	high	control	30°C	10.45	0.22	13
Sweden	high	16 days	20°C	10.38	0.22	13
Sweden	high	16 days	30°C	11.40	0.24	12
Sweden	high	complete F0	20°C	10.17	0.24	12
Sweden	high	complete F0	30°C	11.13	0.22	13
Sweden	low	control	20°C	8.35	0.24	11
Sweden	low	control	30°C	9.48	0.23	12
Sweden	low	16 days	20°C	8.70	0.23	12
Sweden	low	16 days	30°C	9.49	0.23	12
Sweden	low	complete F0	20°C	9.32	0.22	13
Sweden	low	complete F0	30°C	9.36	0.24	11

PO activity

France	high	control	20°C	15.02	0.70	23
France	high	control	30°C	10.87	0.76	21
France	high	16 days	20°C	11.71	0.74	21
France	high	16 days	30°C	10.12	0.76	20
France	high	complete F0	20°C	10.53	0.74	21
France	high	complete F0	30°C	9.06	0.76	21
France	low	control	20°C	11.88	0.71	23
France	low	control	30°C	11.22	0.77	21
France	low	16 days	20°C	11.30	0.75	21
France	low	16 days	30°C	10.53	0.77	21
France	low	complete F0	20°C	10.74	0.75	21
France	low	complete F0	30°C	7.96	0.75	22
Sweden	high	control	20°C	20.88	0.88	16
Sweden	high	control	30°C	18.07	0.79	19
Sweden	high	16 days	20°C	18.47	0.85	18
Sweden	high	16 days	30°C	16.61	0.86	16
Sweden	high	complete F0	20°C	15.18	0.89	13
Sweden	high	complete F0	30°C	14.93	0.82	15
Sweden	low	control	20°C	15.78	0.86	17
Sweden	low	control	30°C	18.65	0.82	17
Sweden	low	16 days	20°C	12.00	0.89	16
Sweden	low	16 days	30°C	10.61	0.88	15
Sweden	low	complete F0	20°C	10.76	0.91	14
Sweden	low	complete F0	30°C	9.52	0.82	17

720

721

722